



**EDMI** Microsystems and Microelectronics

**MICRO-614:** Electrochemical Nano-Bio-Sensing  
and Bio/CMOS interfaces

# Lecture #13

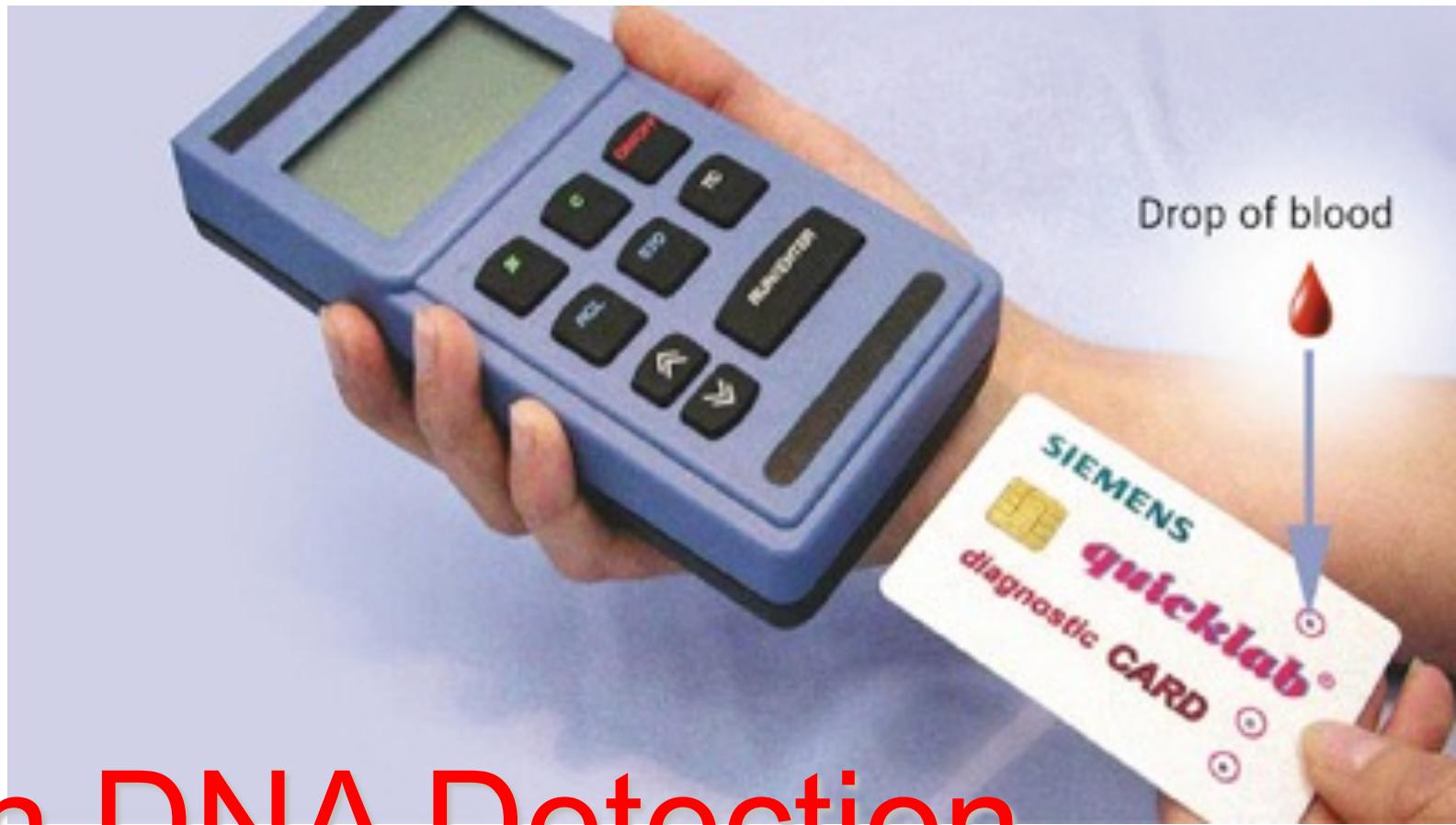
## CMOS interfaces for DNA Detection

# Lecture Outline

(Book Bio/CMOS: Chapter 13

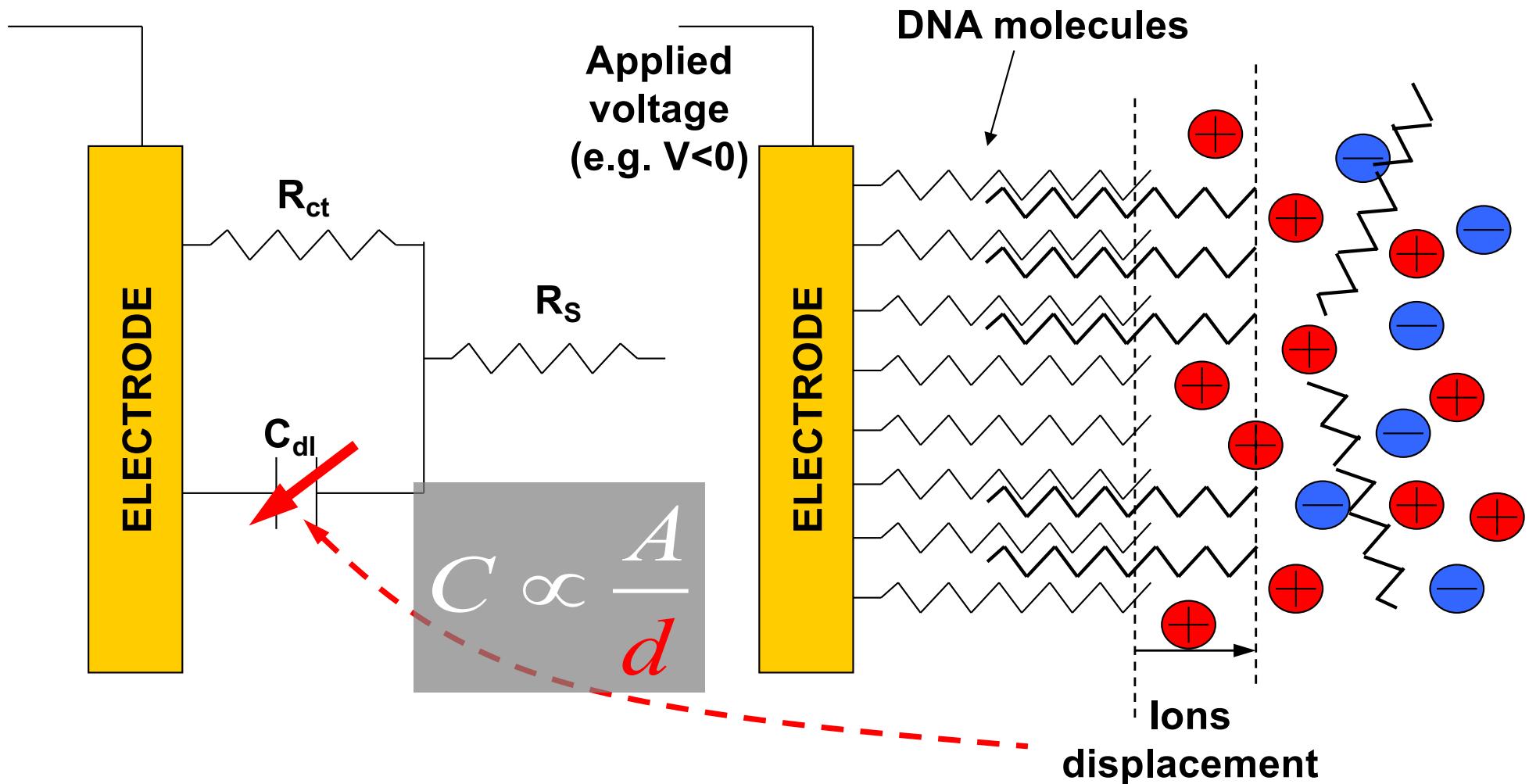
- CMOS for capacitance detection
- Charge-Based Capacitance Measurement (CBCM) Method
- Frequency-to-Capacitance Measurement (FTCM) Method

# CMOS architectures for VLSI



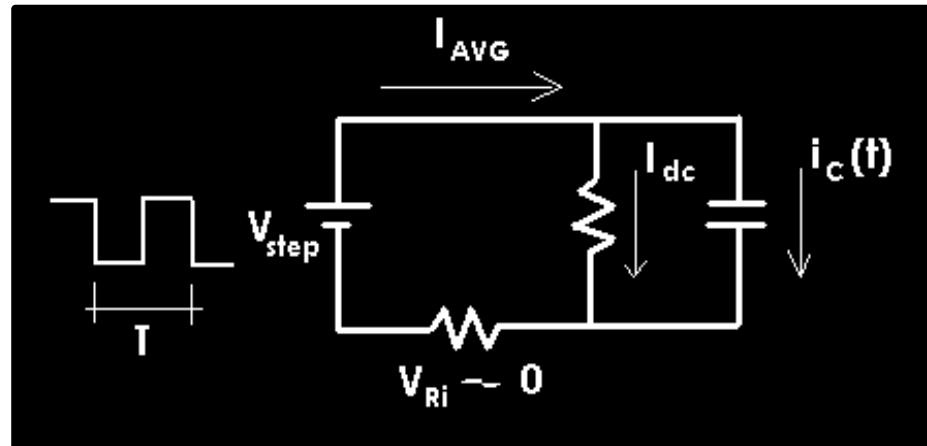
in DNA Detection

# The Capacitance DNA Detection



Unlabeled ssDNA may be detected with capacitance measurements as due to charge displacement

# Current Based Capacitance Measurement (CBCM)



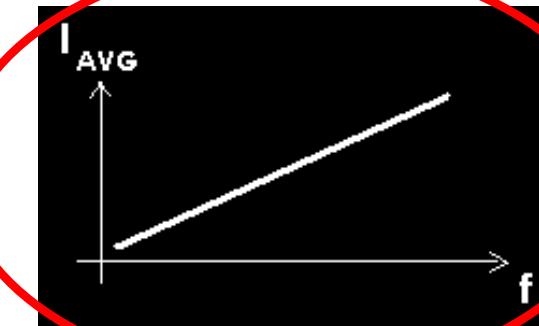
$$i(t) = I_{dc} + i_C(t)$$

Frequency

$$I_{AVG} = \frac{I_{dc}}{2} + \frac{1}{T} \int_0^{T/2} i_C(t) dt$$

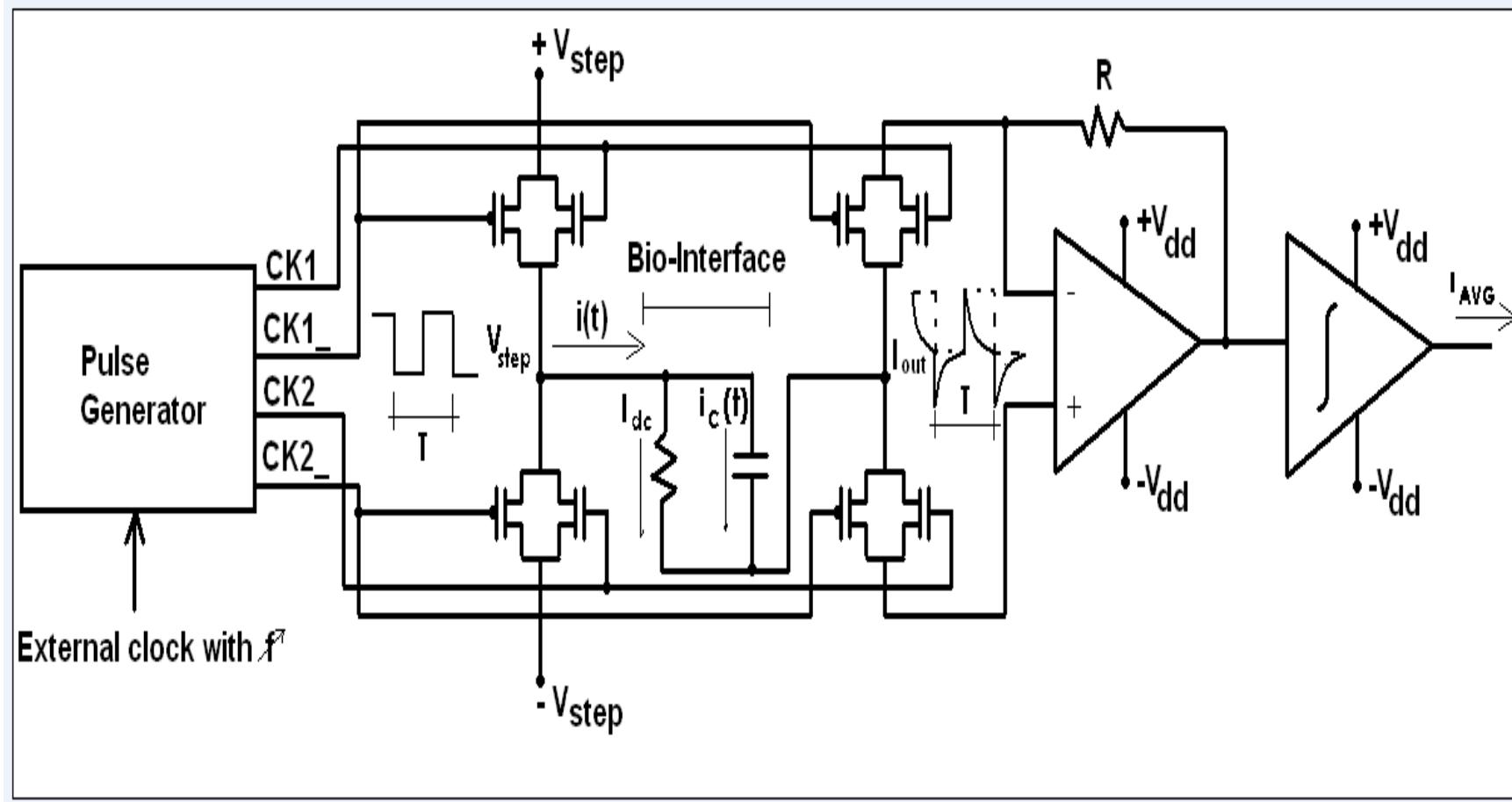
$$I_{AVG} = \frac{I_{dc}}{2} + C V_{step} f$$

THE CAPACITANCE !



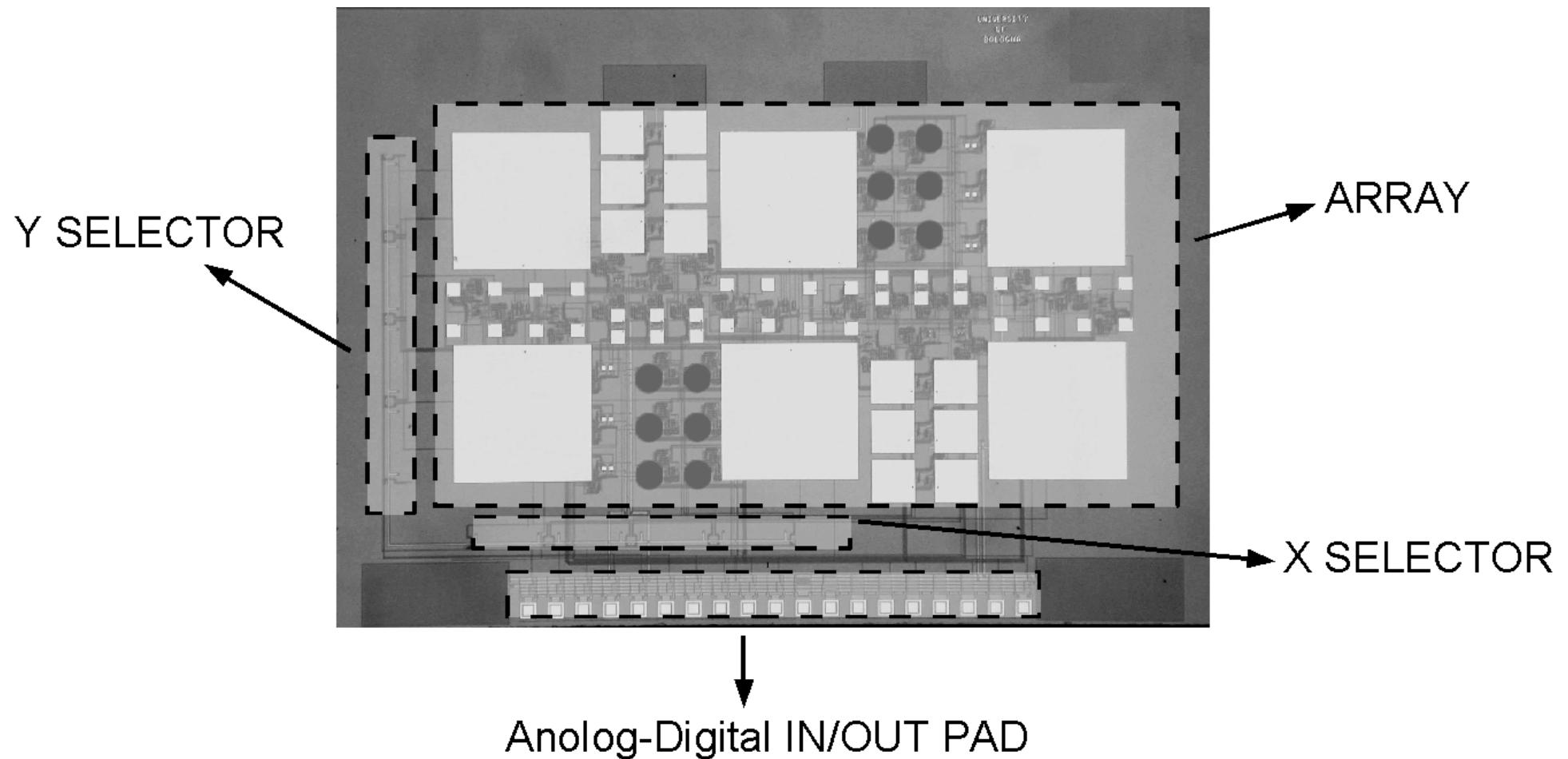
Method for a precise Capacitance measurement

# CMOS for CBCM detection



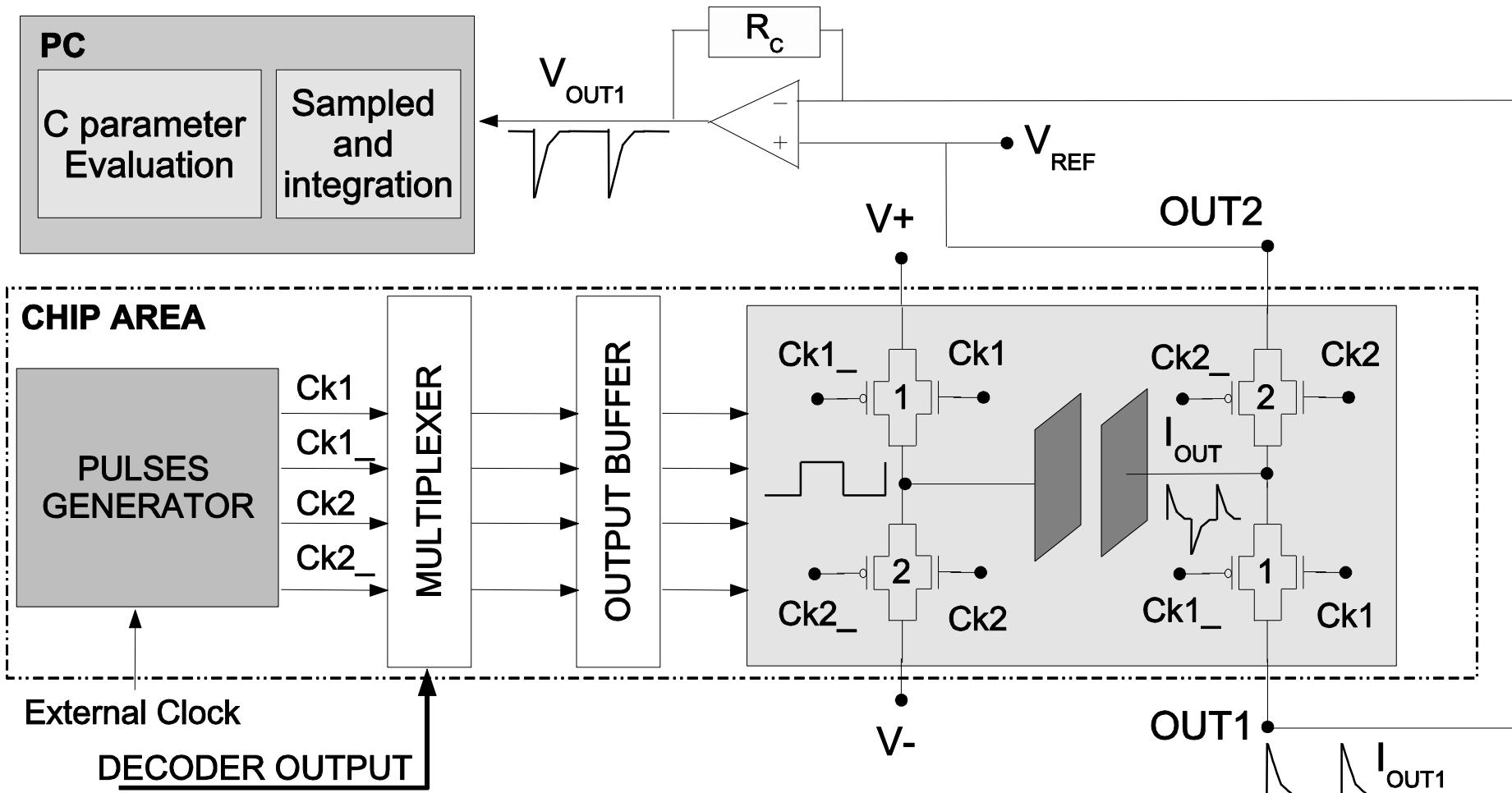
The circuit assures the square signal generator, an inverters, and an integrator to calculate the average current

# *The Chip Electrodes Layout*



# *The VLSI Implementation of the Chip (CBCM method)*

**(CBCM = Charge Base Capacitance Mode)**



# The problem of overlapping signals

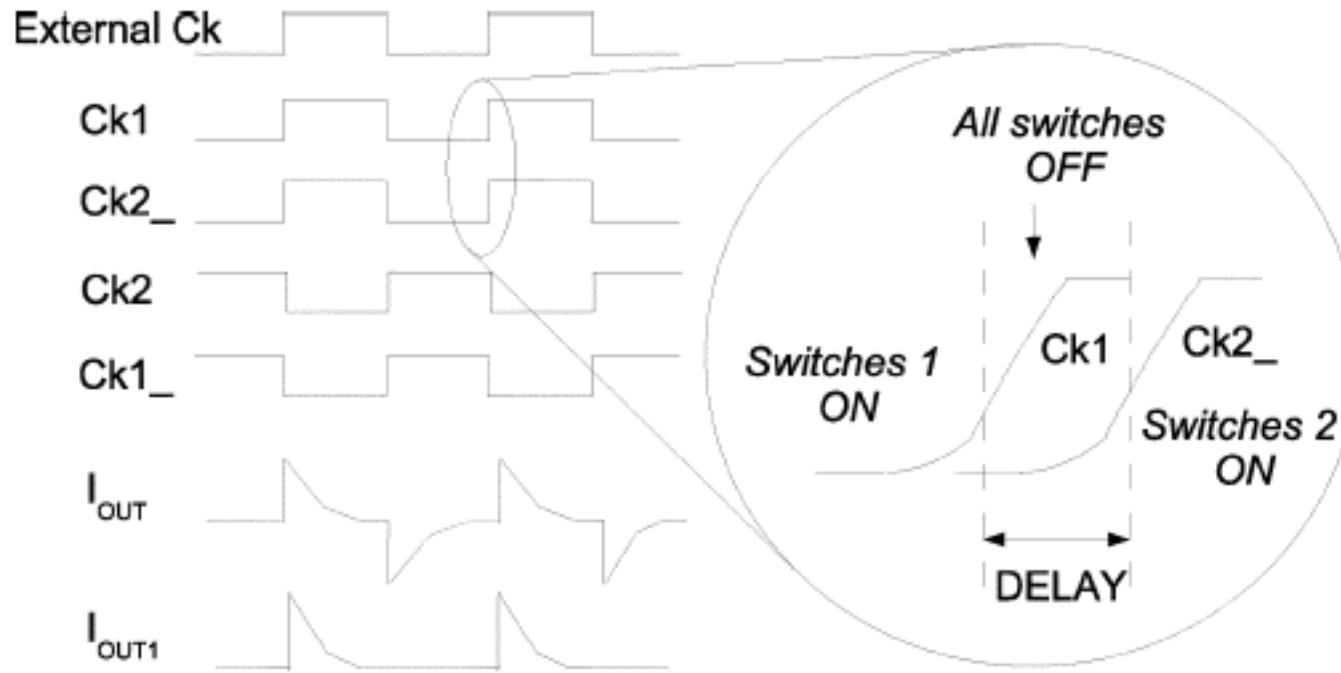


Fig. 4. Schematic representation of the signals flow used in the experiments.

Ck and Ck<sub>\_</sub> signals need to be not-overlapping in order to assure the correct square signal generation

# The circuit solution

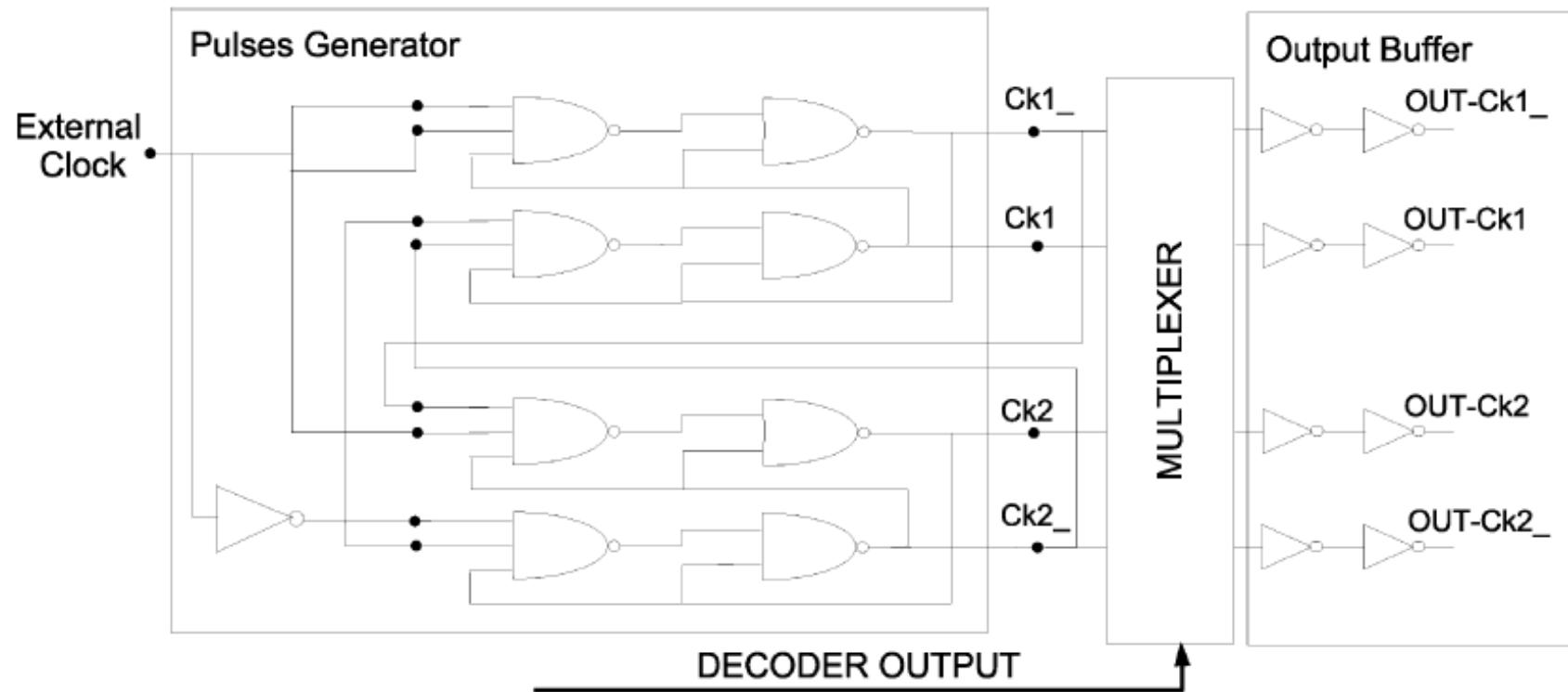


Fig. 5. Schematic plot of the block used to generate not overlapping clock signals.

A simple logical circuit and a digital multiplexer assures not-overlapping Ck and Ck<sub>\_</sub> signals

# The Measurements Set-up

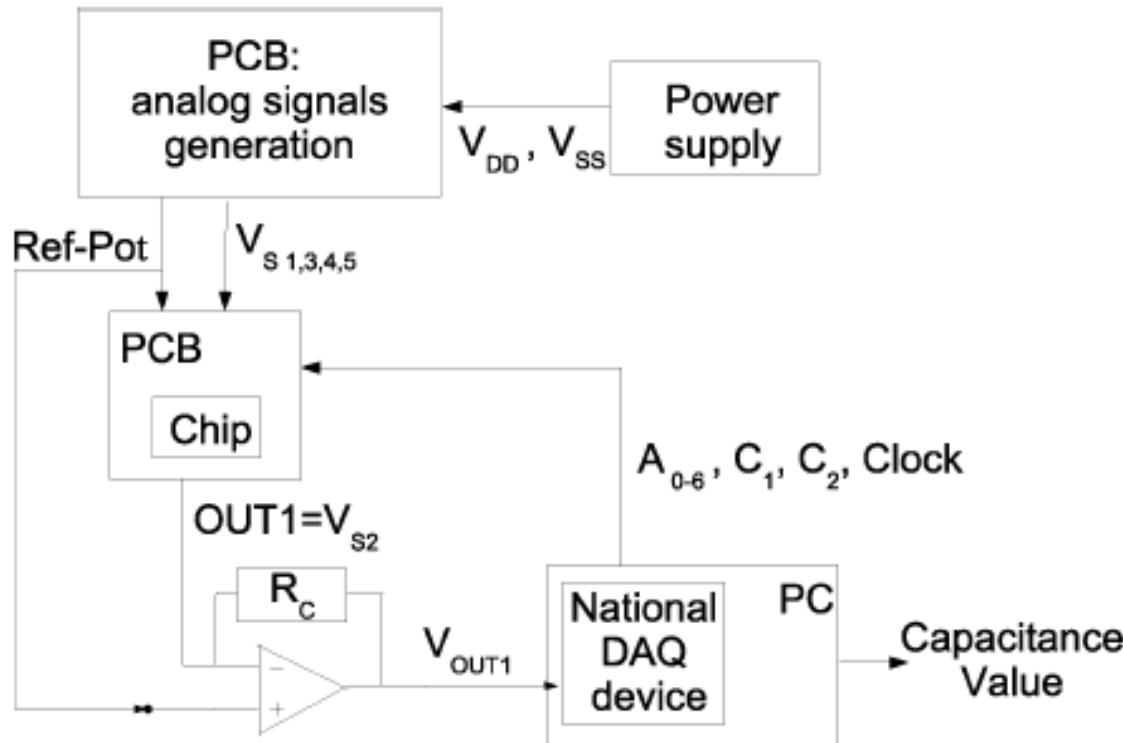
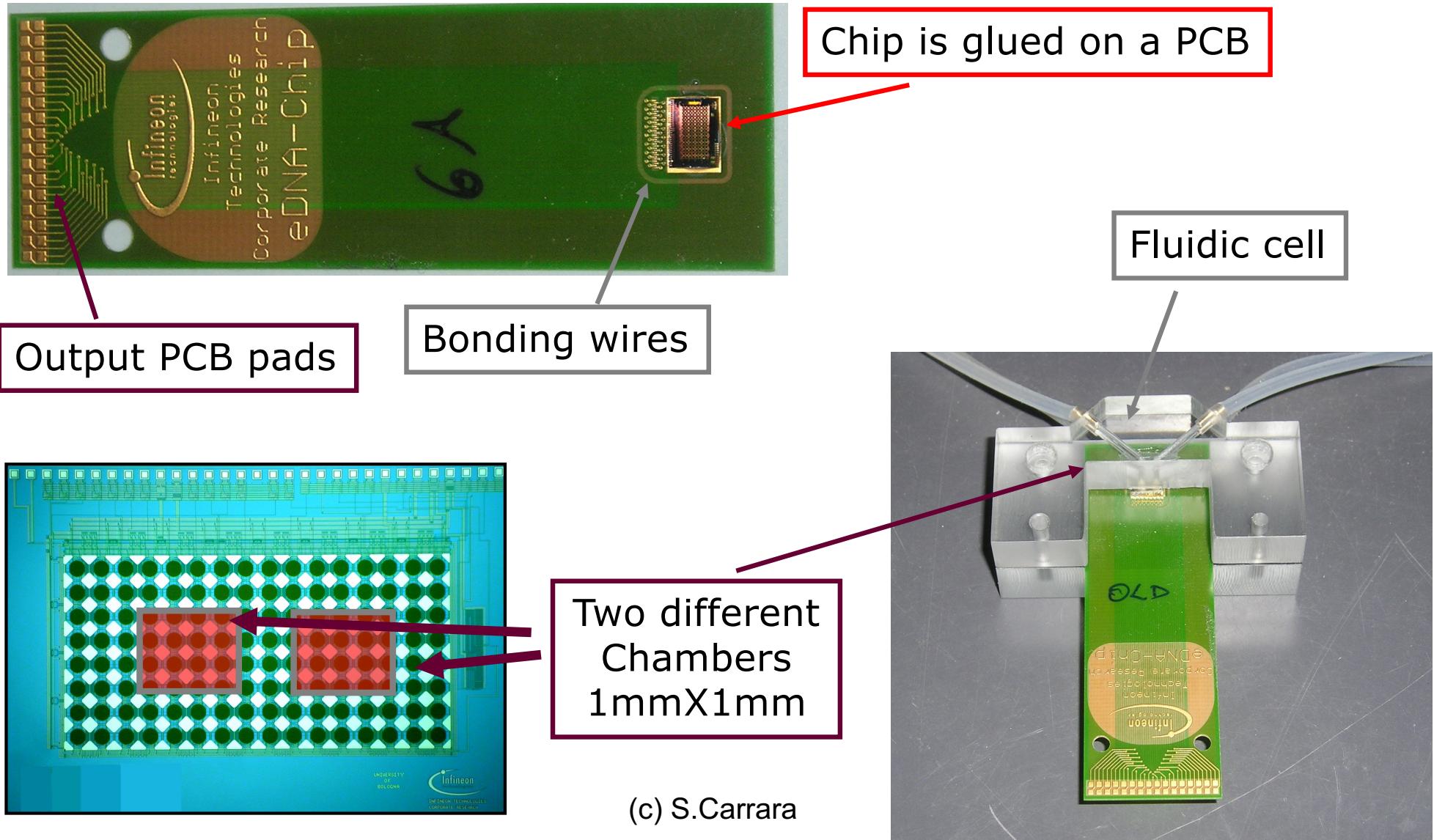


Fig. 8. Schematic representation of the measurement setup.

The Chip has been mounted onto a PCB for PC remote control and testing

# Liquid Measurement set-up



# DNA detection in CBCM mode

Large differences in C values

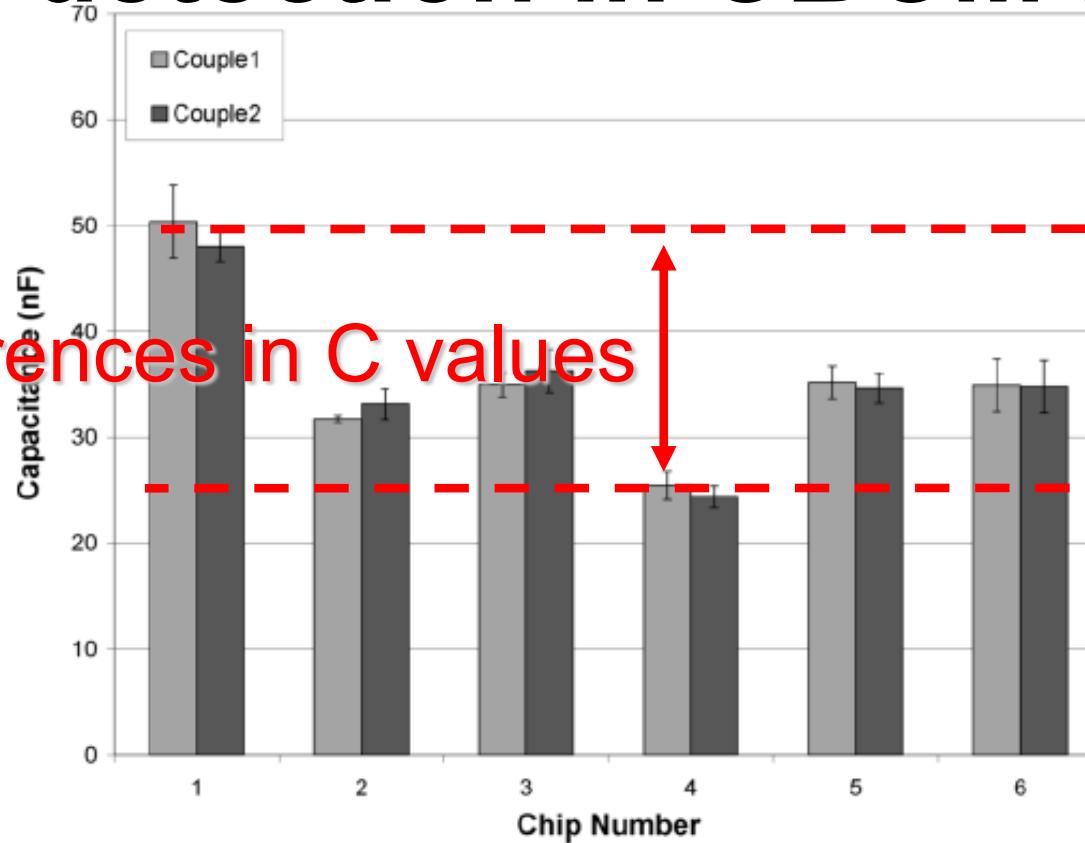


Fig. 10. Capacitance measurements of electrode couples on different chips.

The chip-by-chip reproducibility has been not so high:  
the problem is on the chip electrodes cleaning

# Capacitance vs Frequency

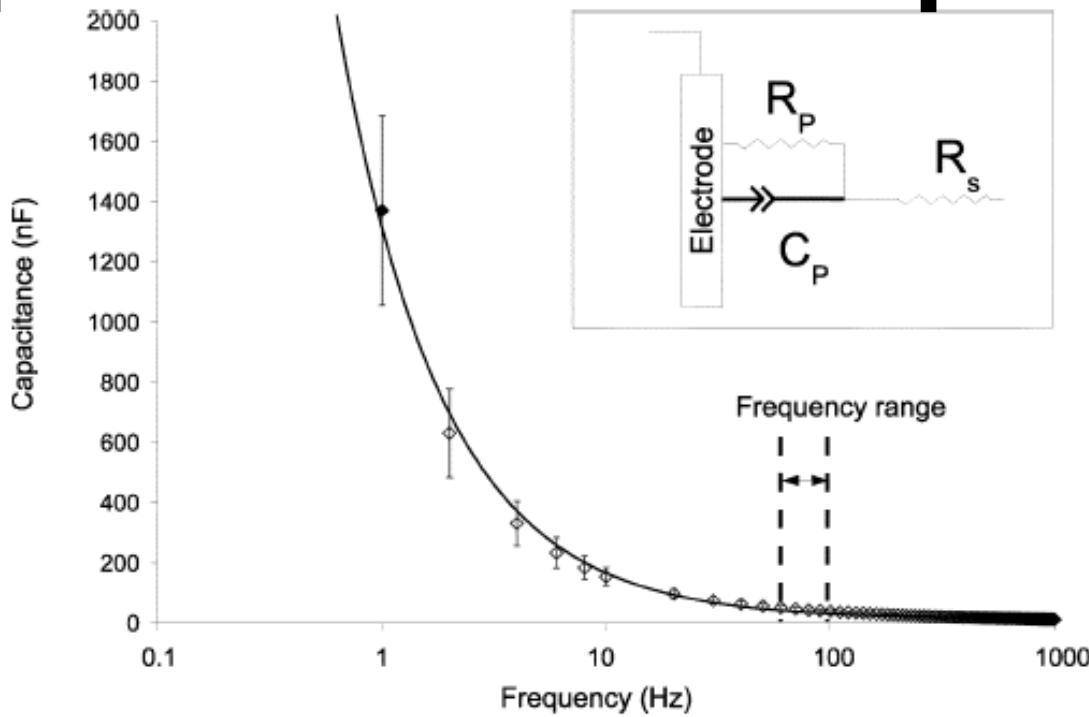
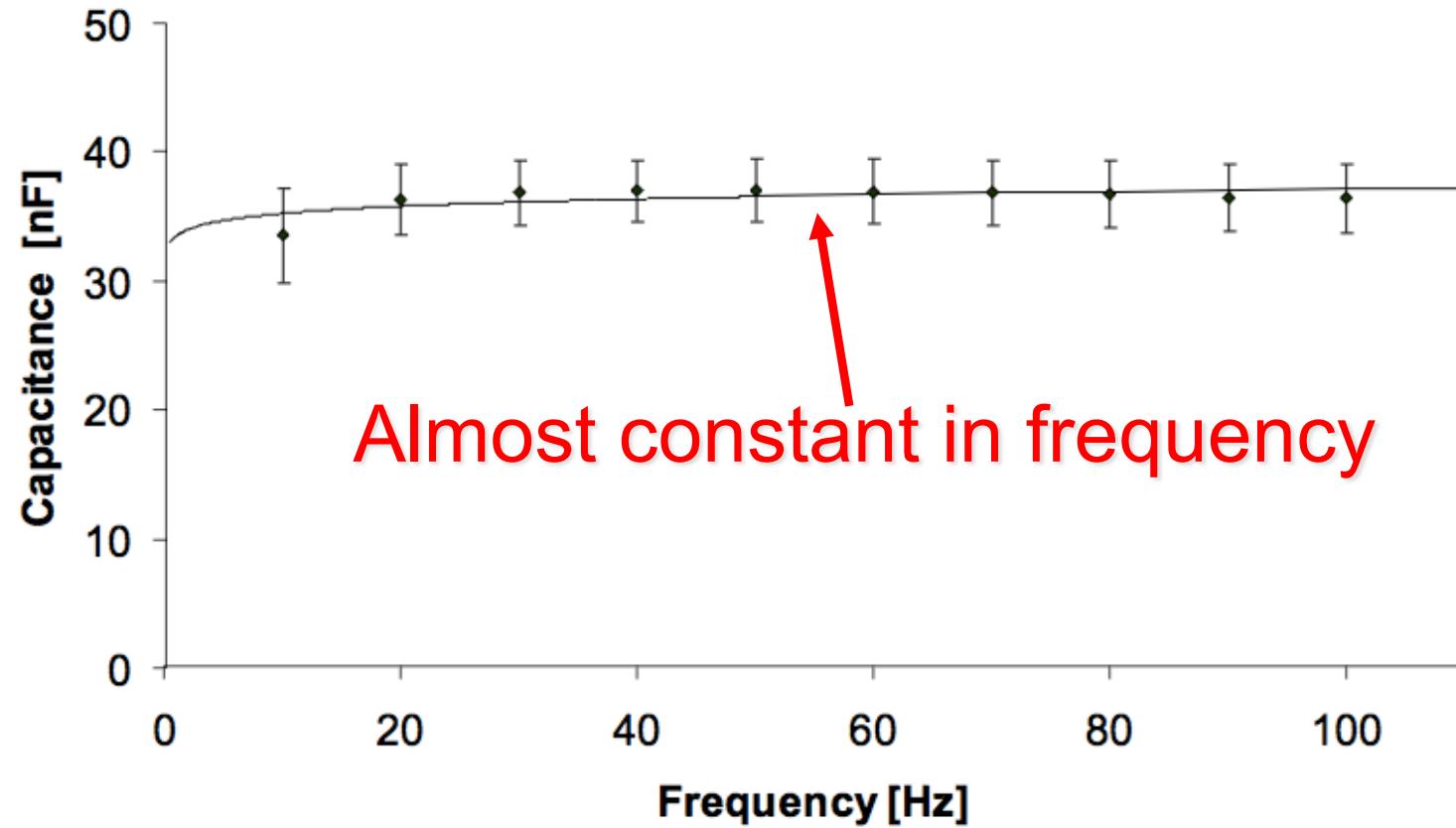


Fig. 9. Measured capacitance versus charge/discharge frequency on clean gold electrodes. The continuous line shows the fitting.

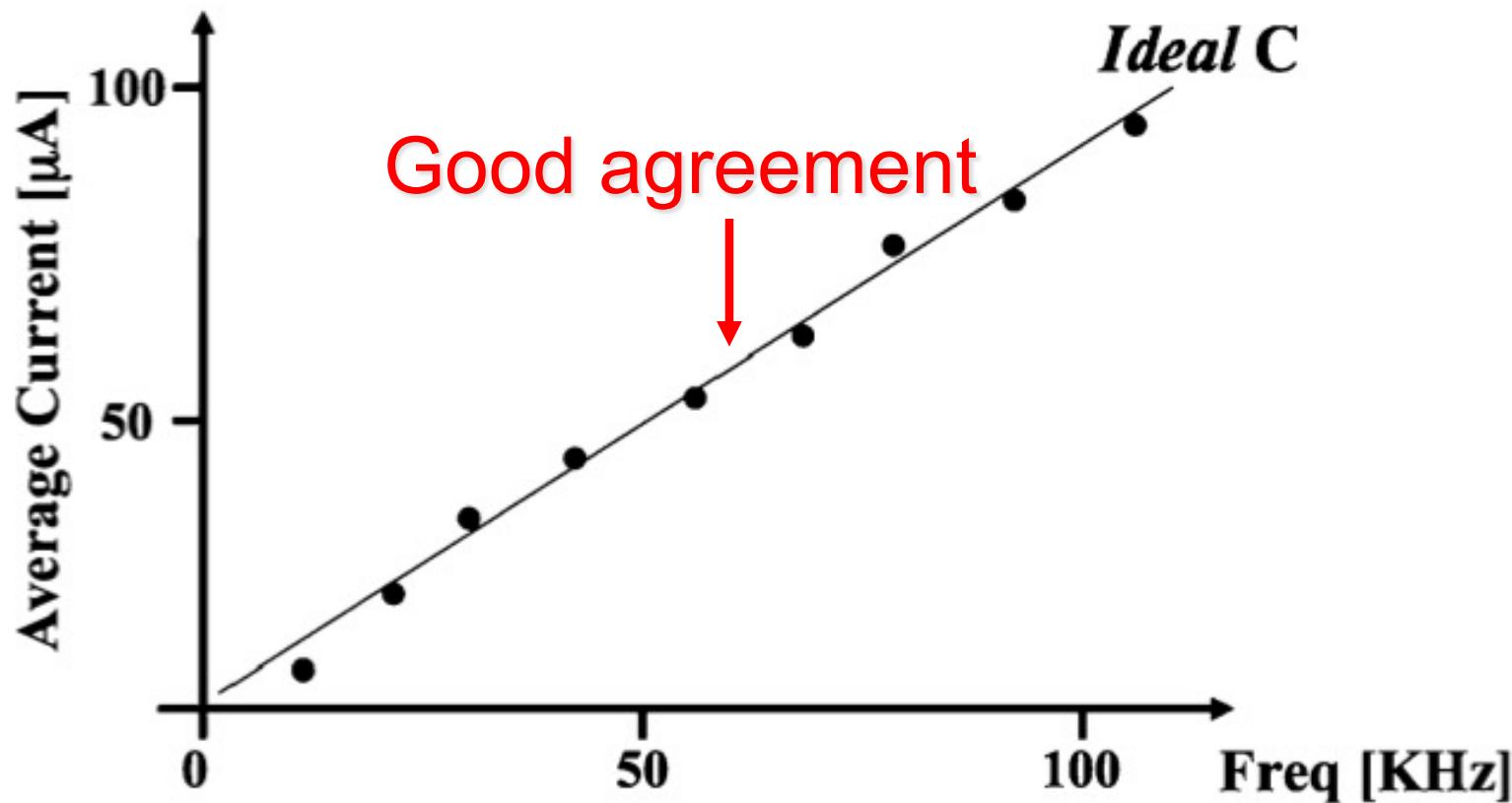
The trends of the measured capacitance vs frequency decrease the accuracy of the measurements in CBCM mode

# Good DNA Layer



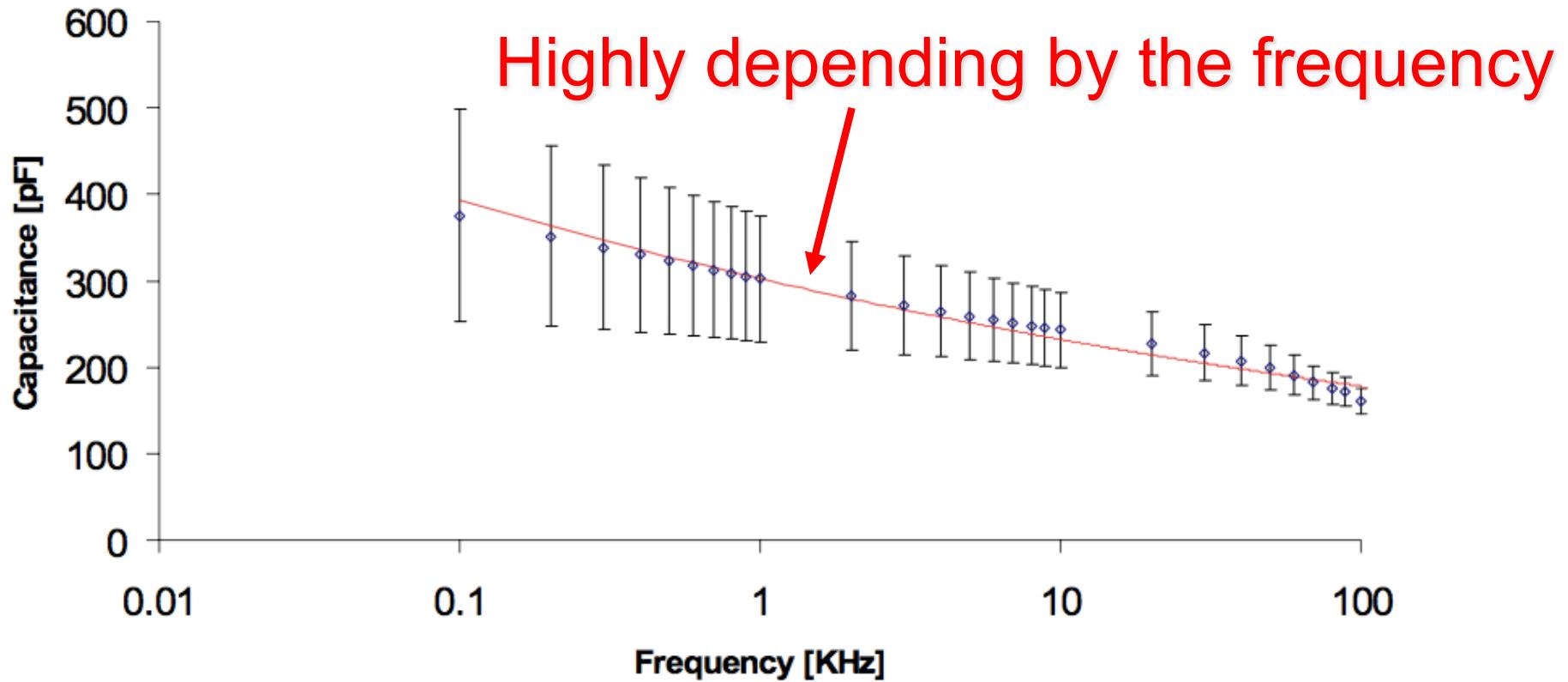
DNA Layer is independent by the frequency thanks to probes immobilized on Ethylene-Glycol Thiols

# CBCM on good DNA Layer



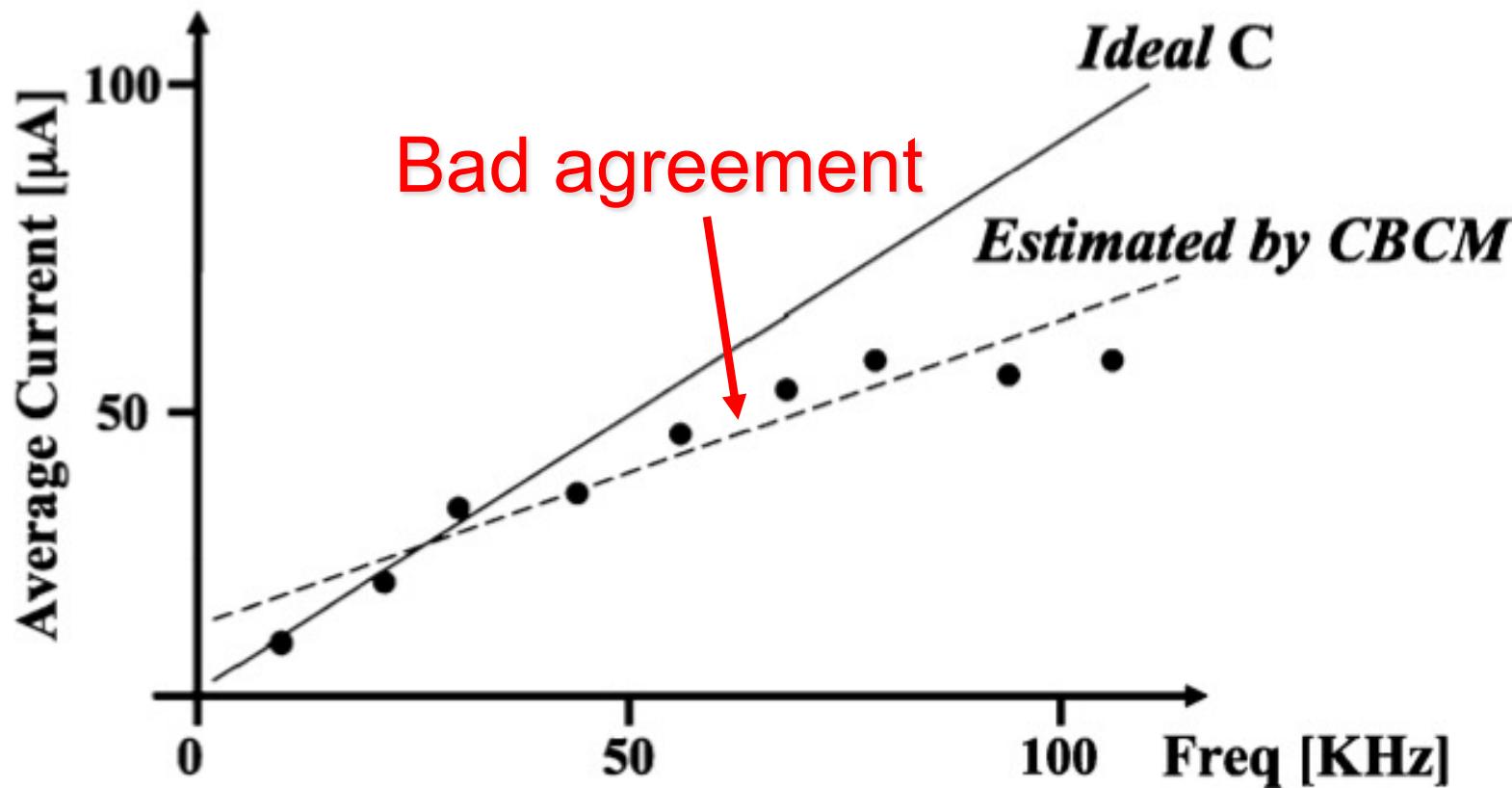
CBCM method on a DNA Layer that is  
independents by the frequency

# Bad DNA Layer



DNA Layer is dependent by the frequency since the monolayer is not extremely well formed

# CBCM on bad DNA Layer



CBCM method on a DNA Layer that  
dependents by the frequency

# DNA detection in CBCM mode

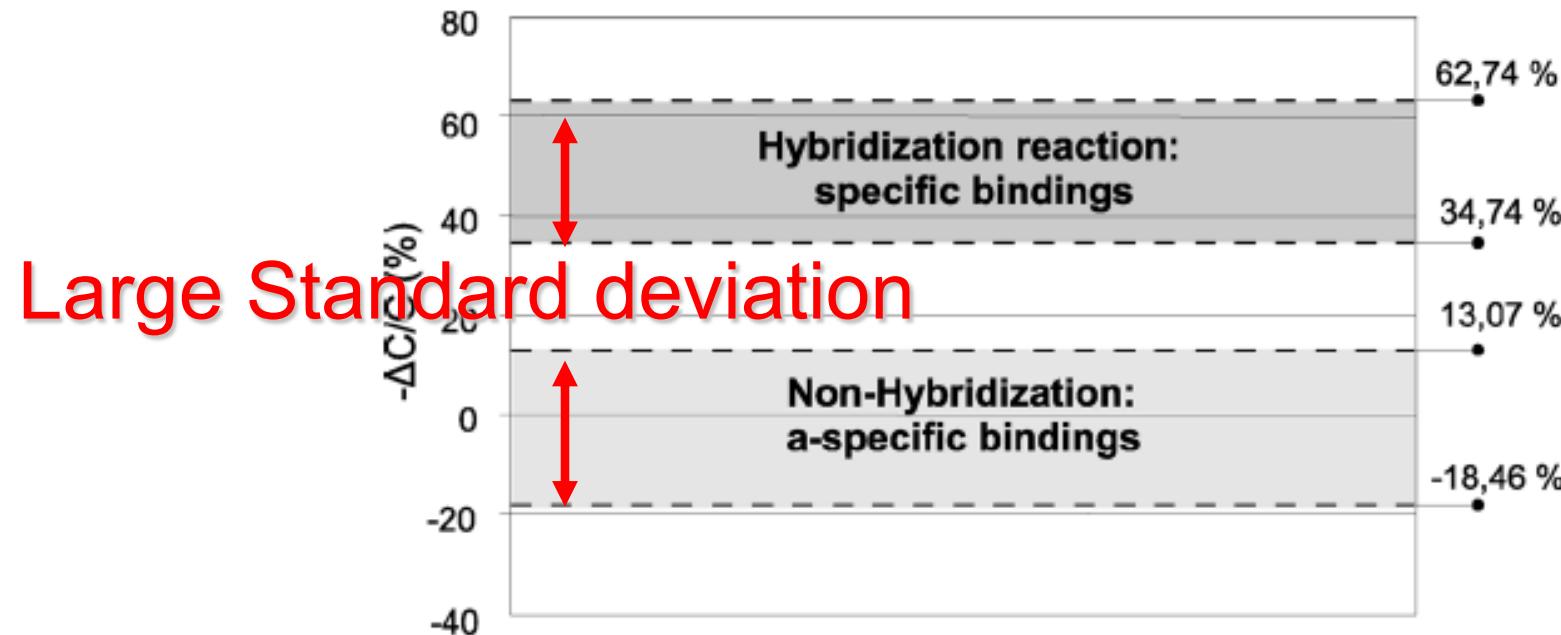
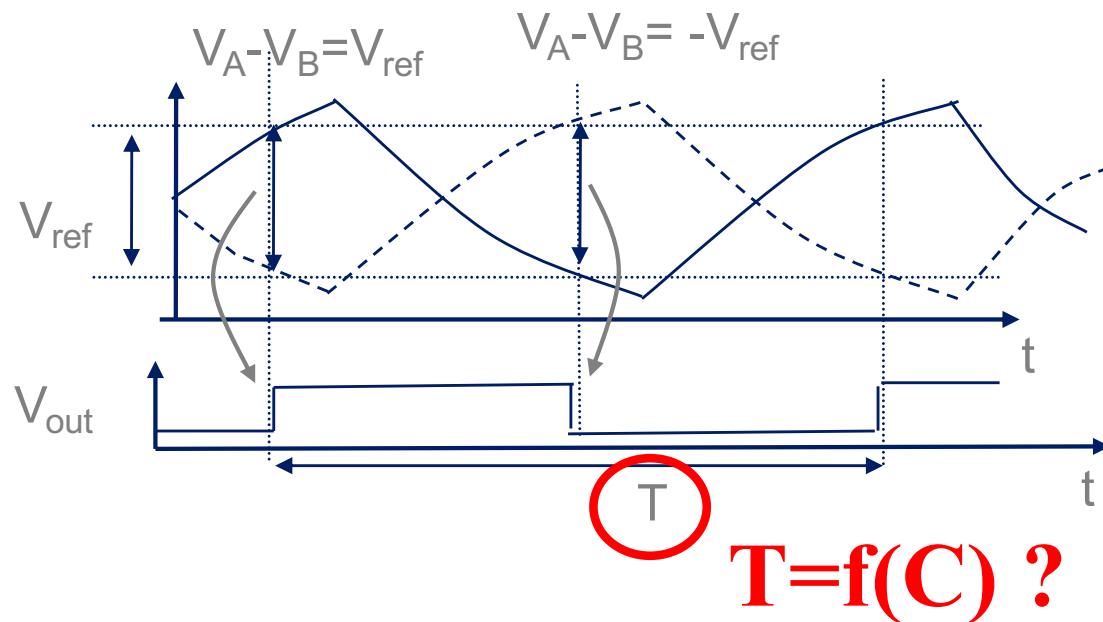
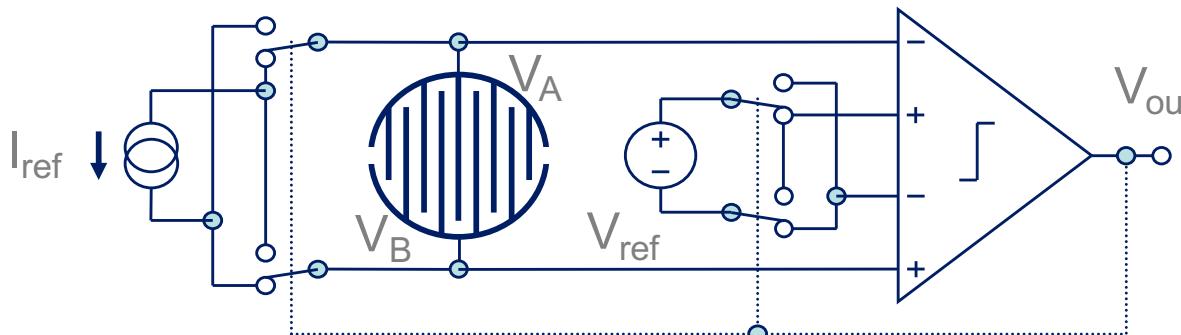


Fig. 12. Capacitance variations due to specific and a-specific bindings (upper and lower bands of measured capacitances, respectively). Positive values indicate capacitance decrease.

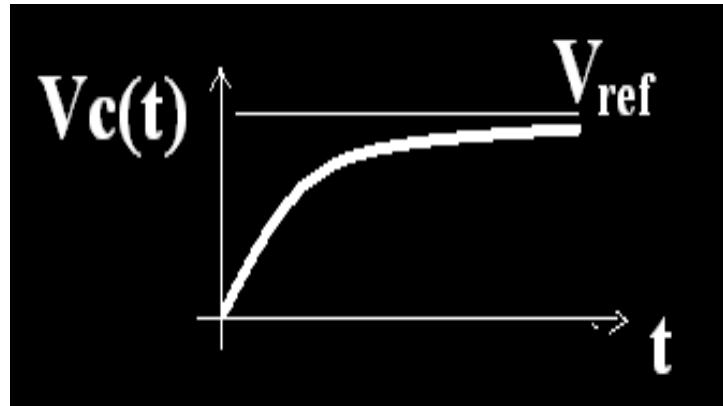
The reproducibility on the same chip-spot is not so high: here the problem is on the nano-scale aperture in the probes surfaces

# Frequency to Capacitance Measurement (FTCM)

**Principle: Frequency To Capacitance Mode**



# Current Based Capacitance Measurement (CBCM)



$$V_c(t) = V_{charge} \left[ 1 - e^{-\frac{t}{RC}} \right]$$

$$V_c\left(\frac{T}{2}\right) = V_{ref} = V_{charge} \left[ 1 - e^{-\frac{T}{2RC}} \right]$$

$$V_c\left(\frac{T}{2}\right) = V_{ref} = RI_{ref} \left[ 1 - e^{-\frac{T}{2RC}} \right]$$

$$\frac{T}{2RC} = -\ln \left[ 1 - \frac{V_{ref}}{RI_{ref}} \right] \rightarrow T = 2RC \ln \left[ 1 - \frac{V_{ref}}{RI_{ref}} \right]^{-1}$$

# The Taylor Series

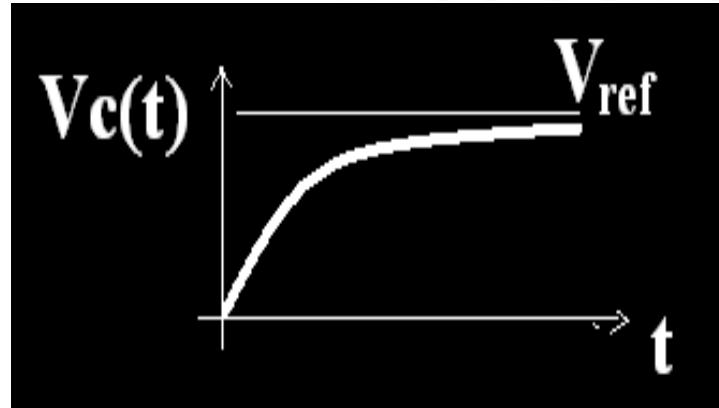
$$f(x) = f(0) + \left\{ \frac{\partial f(x)}{\partial x} \right\} x + \left\{ \frac{\partial^2 f(x)}{\partial x^2} \right\} x^2 + o(3)$$

$$\frac{1}{1-x} = 1 + \left\{ -(-1) \right\} x + o(2) \cong 1 + x$$

$$\ln \left[ \frac{1}{1-x} \right] = \ln [1 + x + o(2)] \cong \ln [1 + x] = 0 + x + o(2) \cong x$$

Linearity by approximation in the right range of values

# Current Based Capacitance Measurement (CBCM)



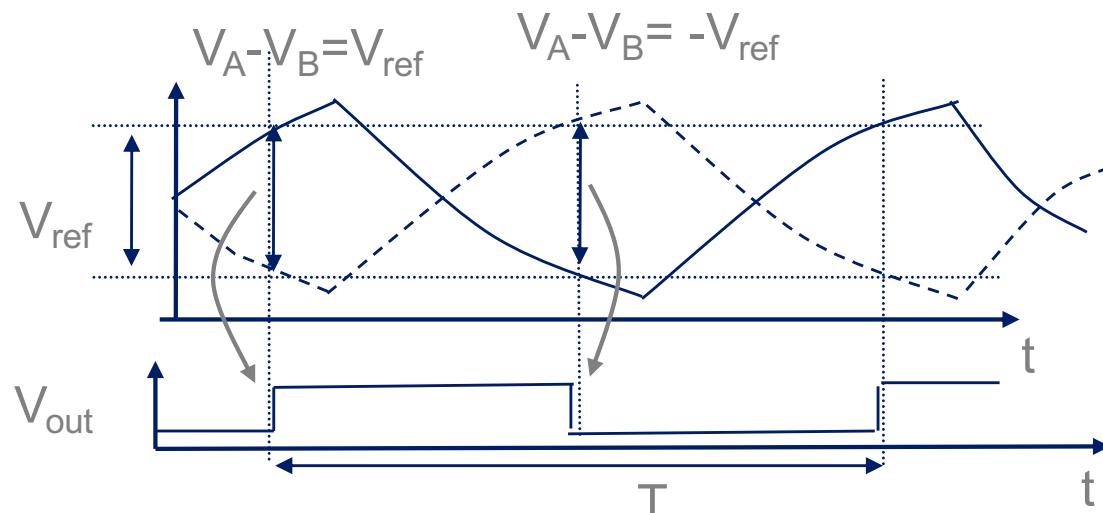
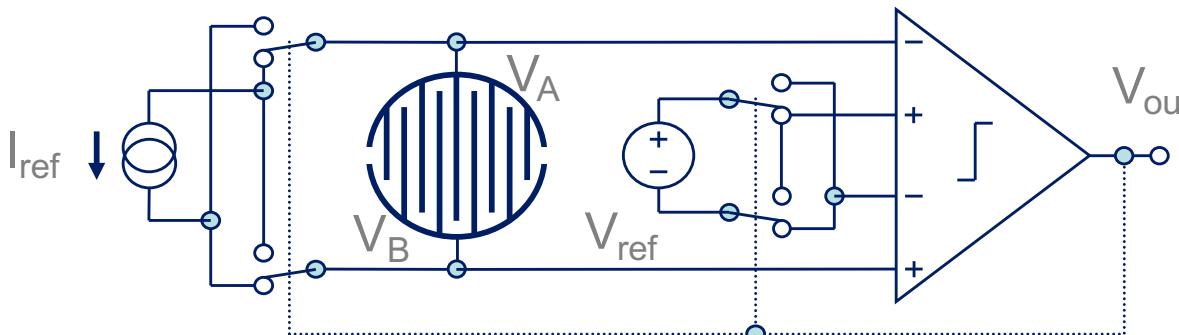
$$T = 2RC \ln \left[ 1 - \frac{V_{ref}}{RI_{ref}} \right]^{-1}$$

$$T = 2RC \ln \left[ \frac{1}{1 - \frac{V_{ref}}{RI_{ref}}} \right] \cong 2RC \ln \left[ 1 + \frac{V_{ref}}{RI_{ref}} \right] \cong \frac{2CV_{ref}}{I_{ref}}$$

Method for the estimation of the Capacitance

# Frequency to Capacitance Measurement (FTCM)

**Principle: Frequency To Capacitance Mode**

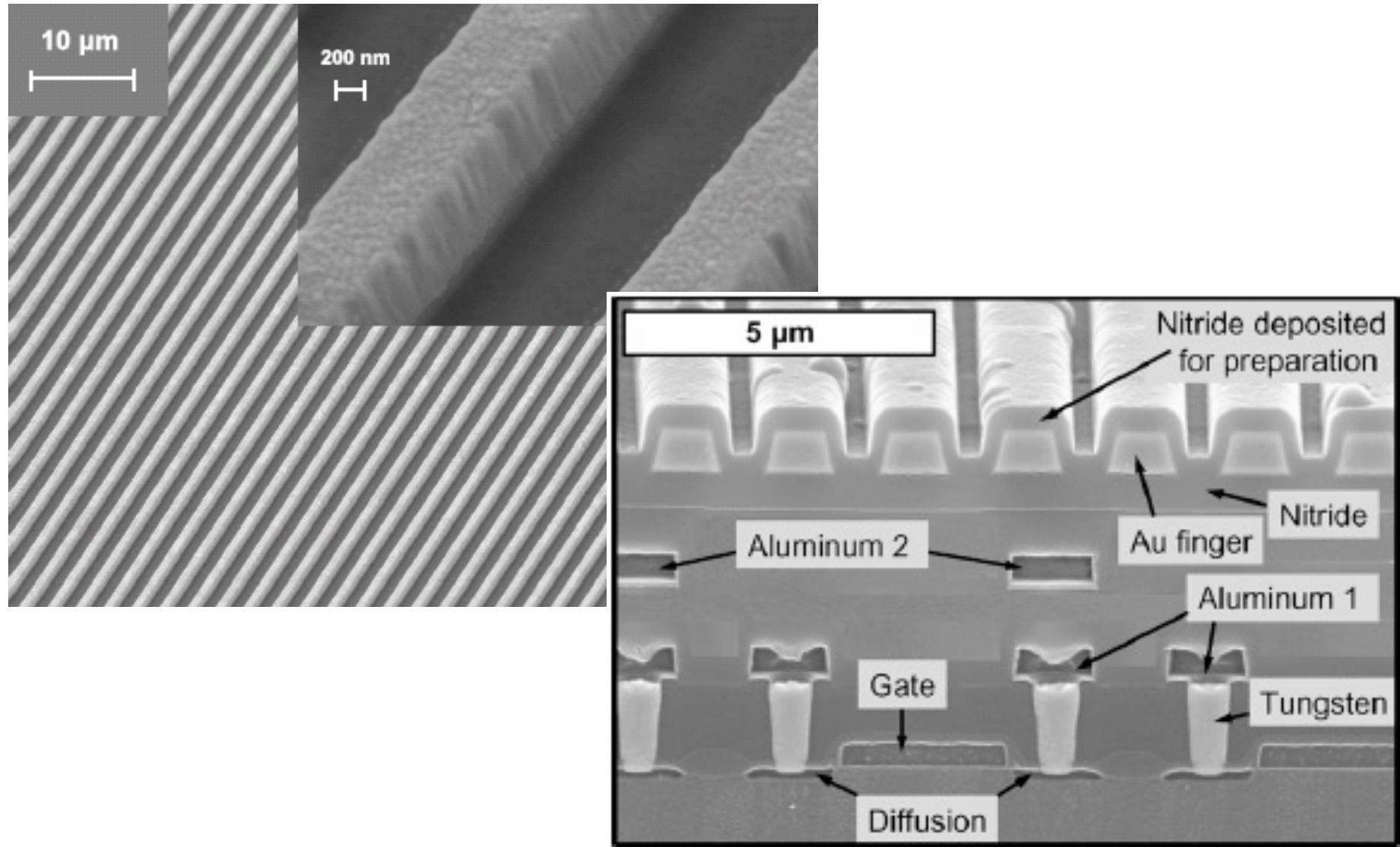


$$T = 2RC \ln \frac{1}{1 - \frac{V_{REF}}{I_{REF}R}}$$

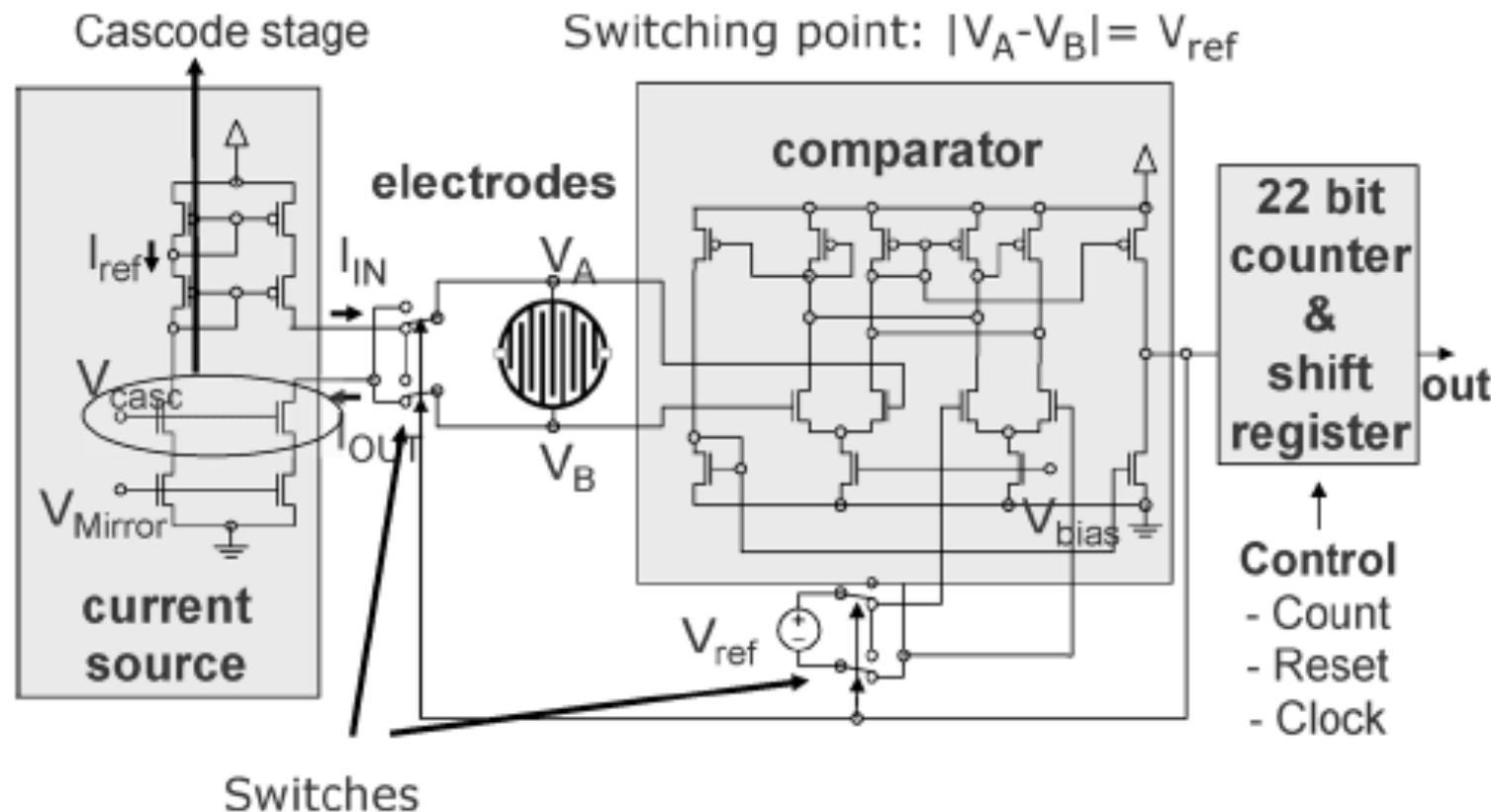
$V_{REF}/I_{REF}R \rightarrow 0$

$$T = \frac{2 \cdot V_{REF} \cdot C}{I_{REF}}$$

# *Electrodes Layout*



# Chip Architecture (FTCM)



# Measurements Set-up

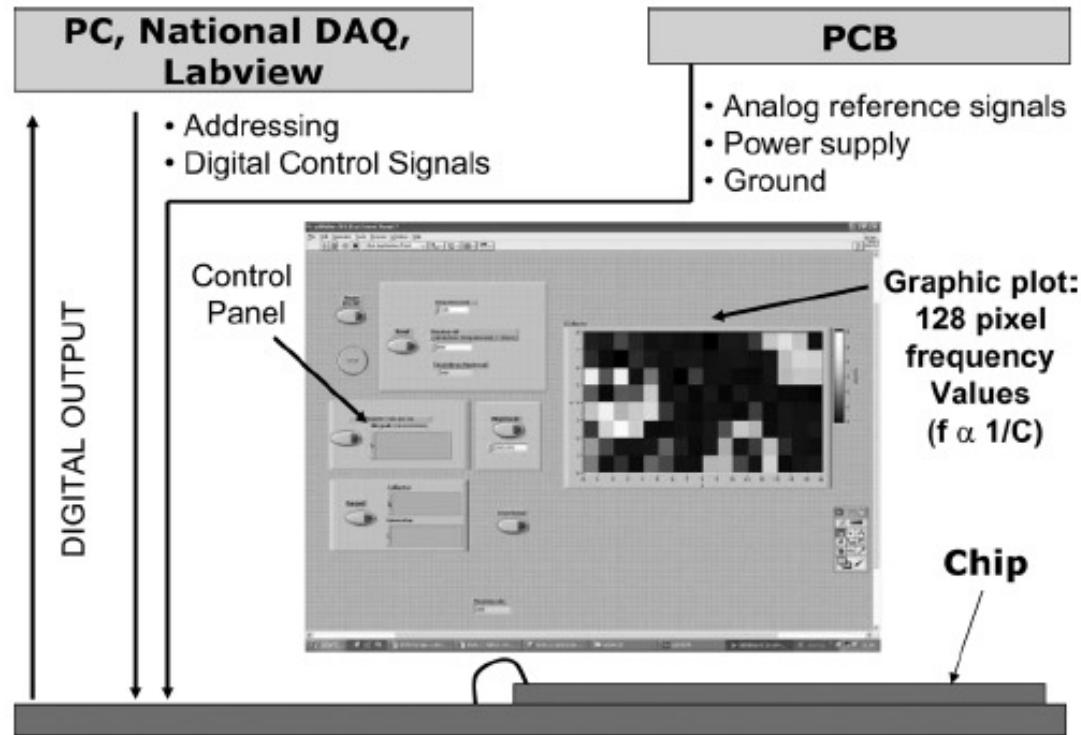


Fig. 9. Schematic representation of the measurement set-up. Voltage reference signals and power supply are generated by circuitry on the PCB. Digital control signals are provided by a PC. The LabView interface manages all the parameters involved in the measurements and shows directly on the screen the measurement results of the whole array.

# Validation Test

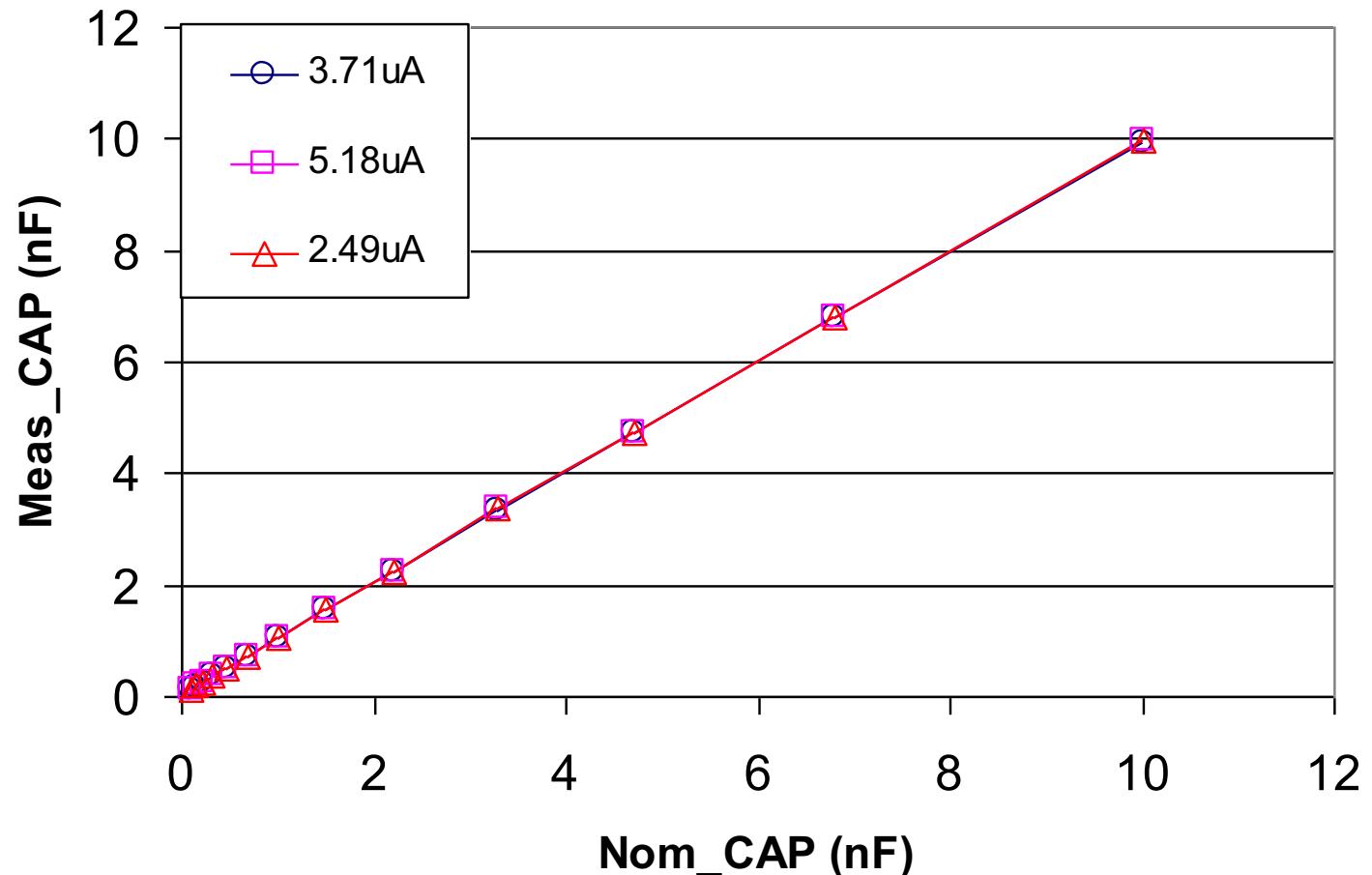
**A test structure has been implemented on chip beside the array to characterize the measurement circuit with discrete test capacitances (10 pF -10 nF)**

**Slope = 0.9837**

**Intercept = 62 pF**

**$\sigma < 0,3 \%$**

*Offset is due to parasitic capacitances of cables*



# Probes property on FTCM mode

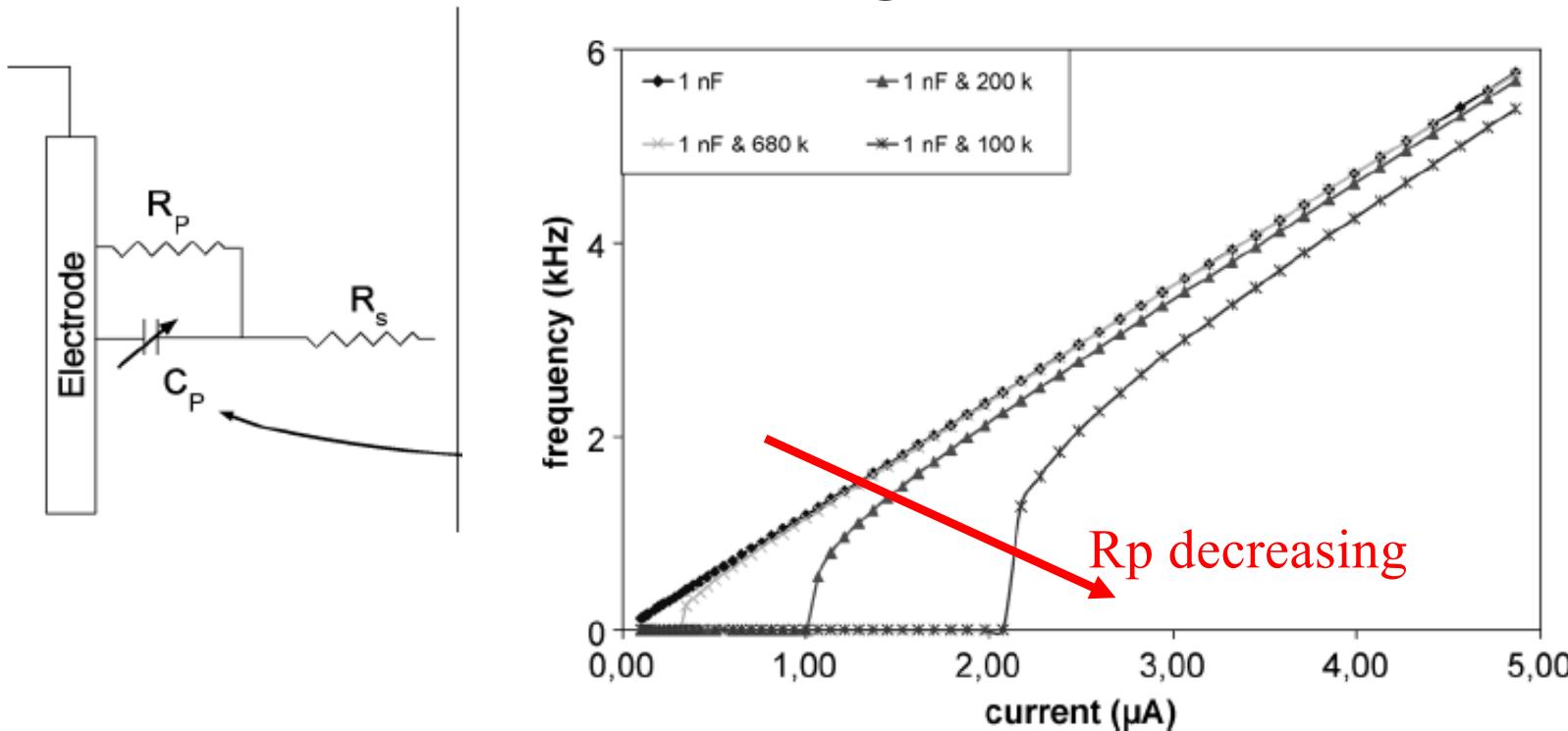
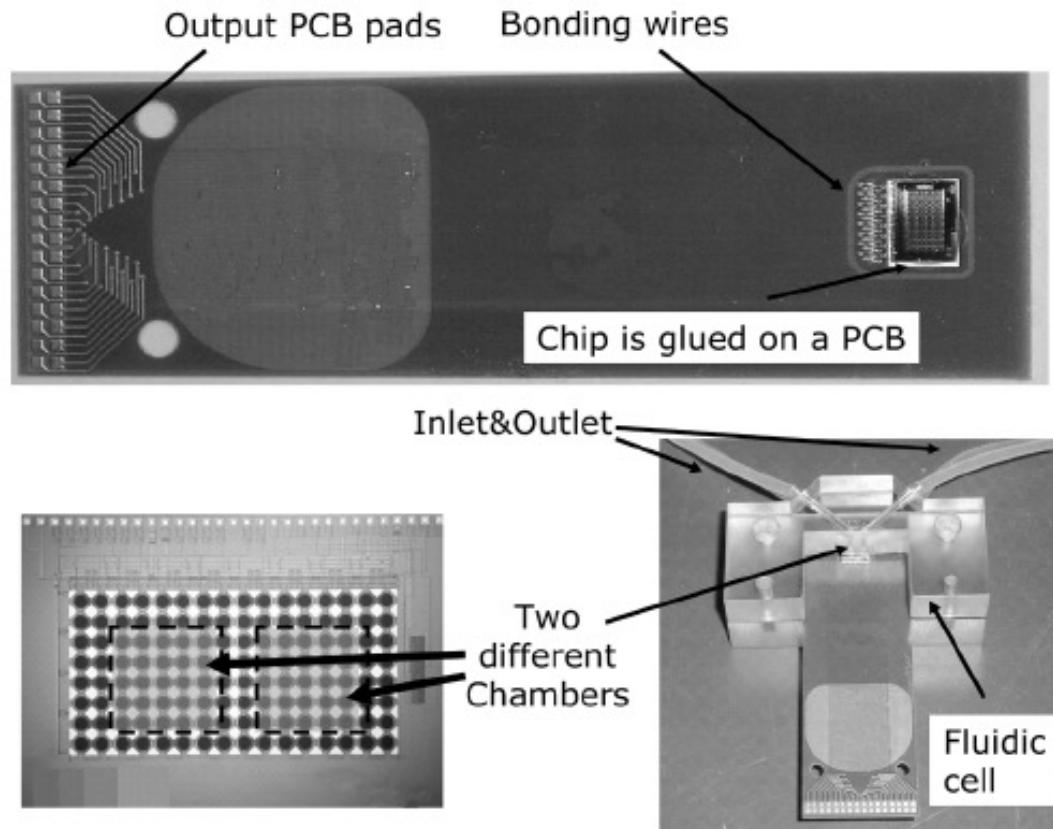


Fig. 12. Frequency versus reference current showing that a significant influence of the parallel resistance on the measurement result occurs only at low current values and at  $R_p$  values lower than  $680\text{ k}\Omega$ .

The linearity between the current and the measured frequency is lost at low current if the CMOS/Bio interface is not a perfect capacitor

# Liquid Measurement set-up



# DNA detection in FTCM mode

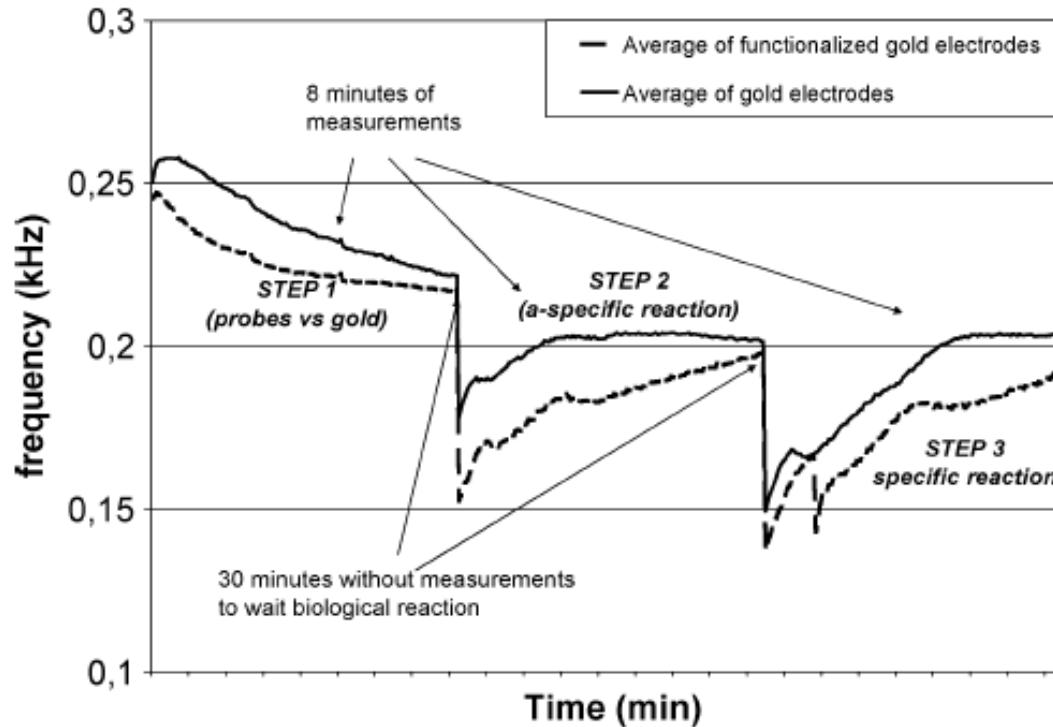


Fig. 13. Frequency changes of the average of reference electrodes (continuous line), and the average of functionalized electrodes (dashed line) show a larger gap after DNA hybridization step considering the stable value reached at the end of the transient.

Time stability on the single chip-spot is poor due to nano-scale aperture in the probes surfaces

# DNA detection in FTCM mode

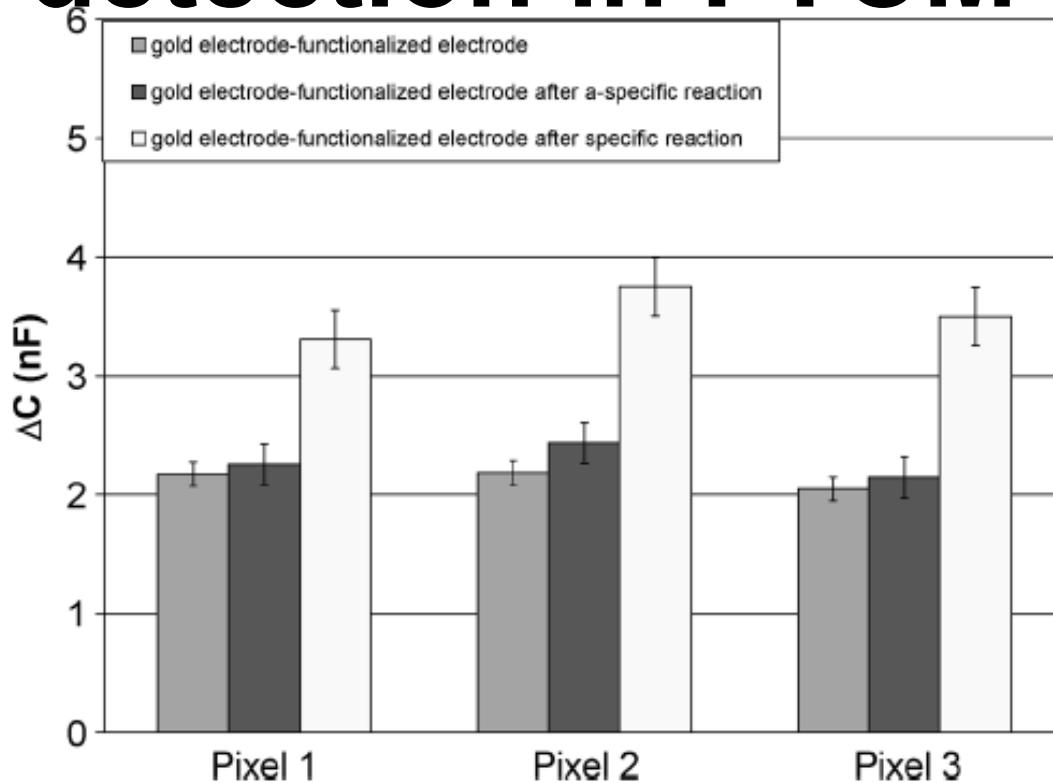
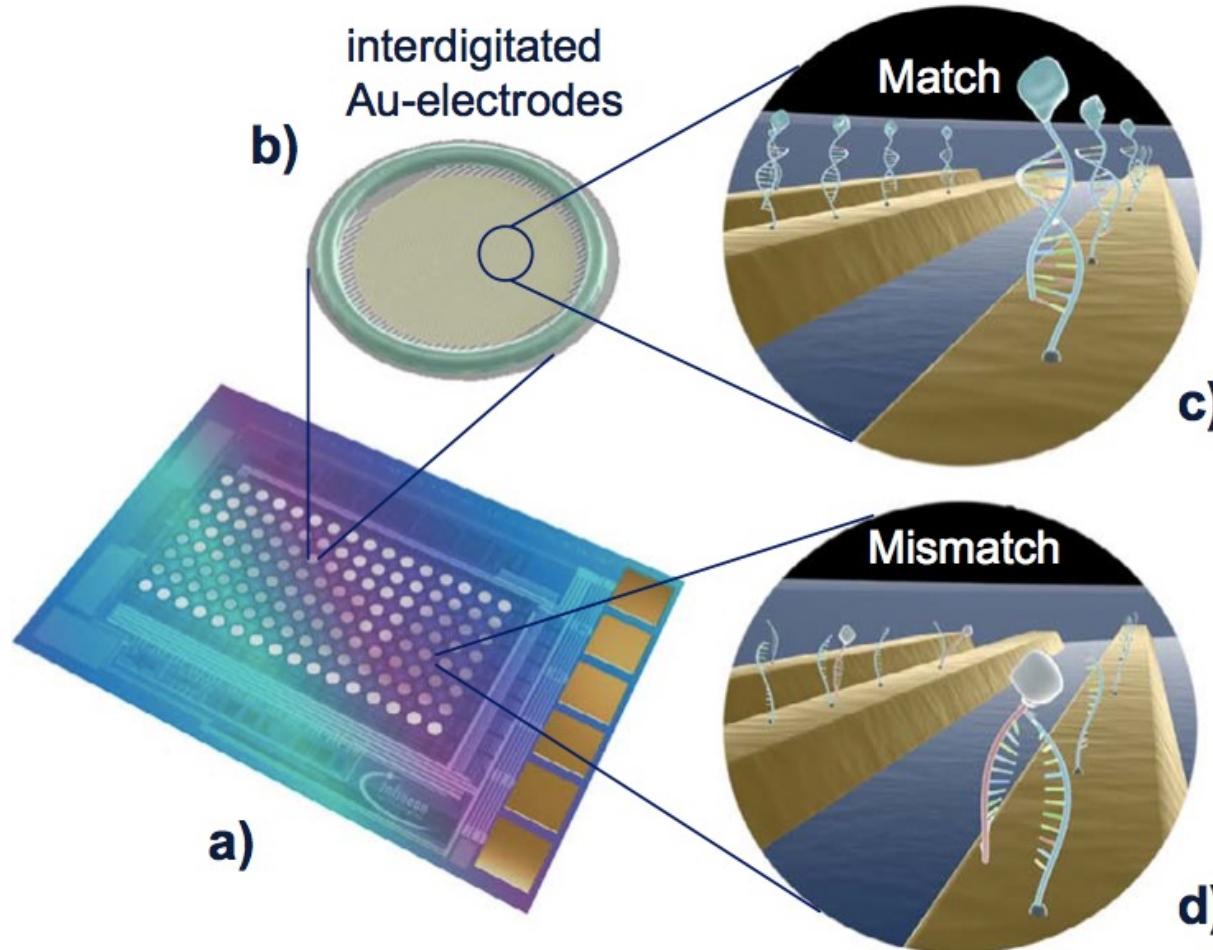


Fig. 14. Typical variations for several pixels among functionalized electrodes and the average value of reference gold electrodes. Capability to distinguish between specific and nonspecific binding is shown for each pixel.

In chip spot-by-spot reproducibility is improved due to better cleaning of the spot gold electrodes

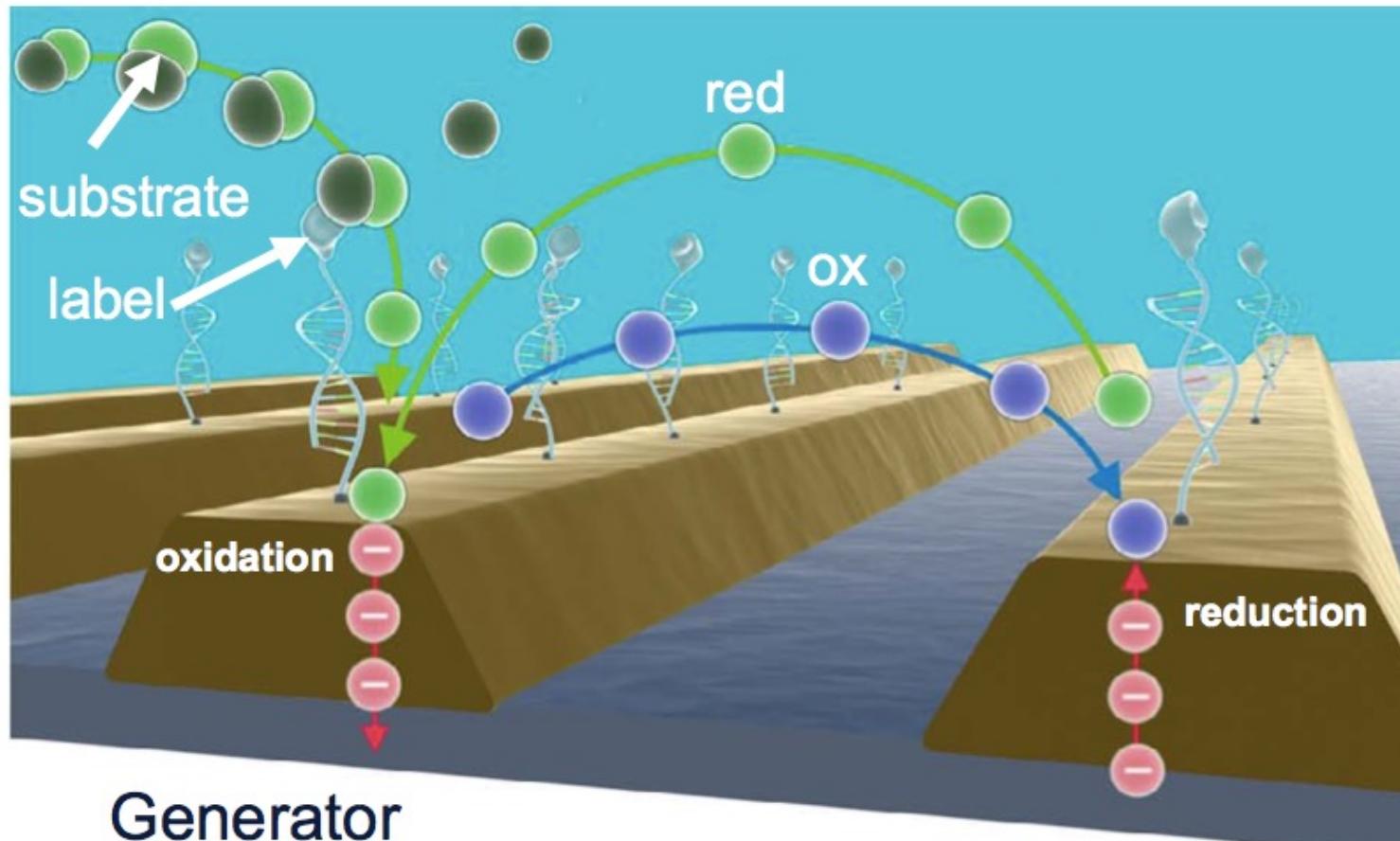
# Amperometric Detection of DNA

Figure by Frey et al, IEEE ISCAS 2015



Electrochemical labels might be used to detect DNA

# Amperometric Detection of DNA

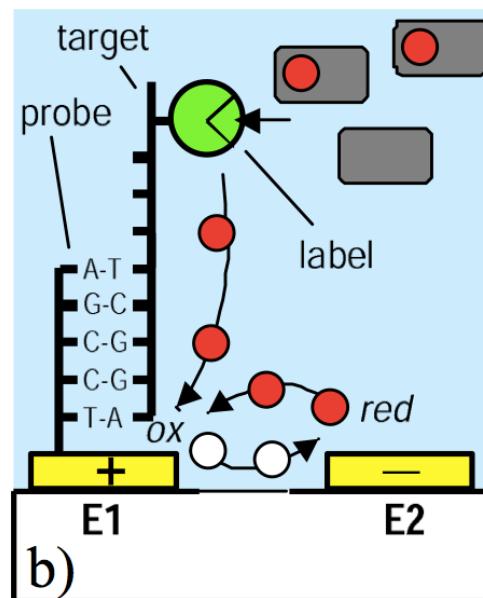
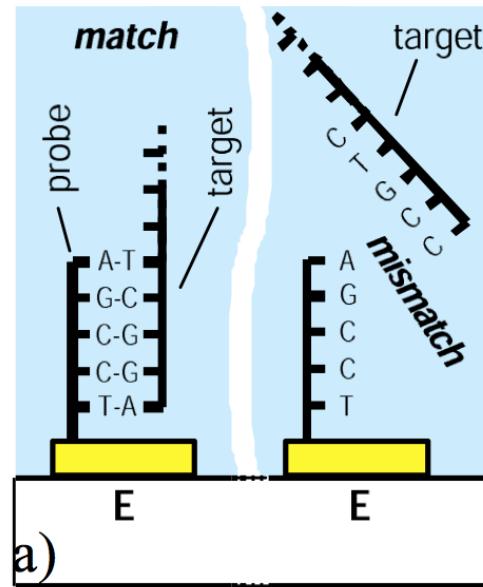


e)



Redox species can be then measured at the electrodes

# Amperometric Detection Principle

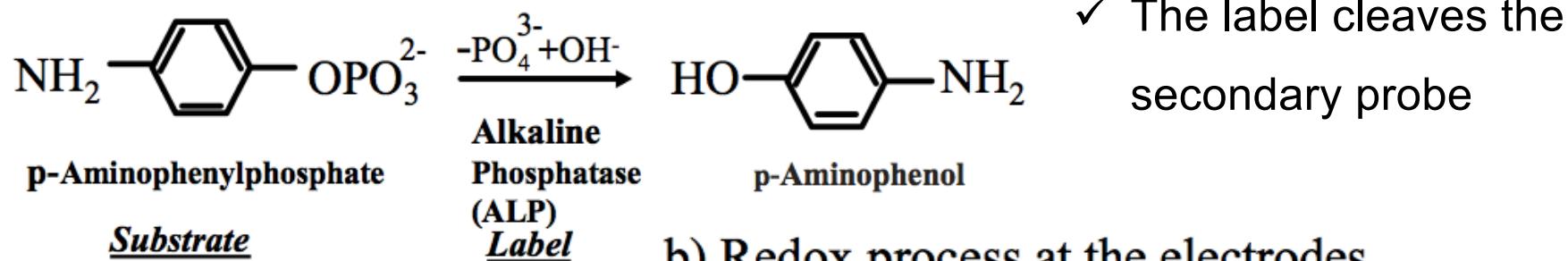


## How it works:

- ✓ First, single stranded DNA molecules (about 20 bases) are immobilized by using a spotting machine on top of the gold electrodes due to gold-thiol coupling.
- ✓ Then, the chip is flooded with an analyte containing labeled target DNA ss: hybridization takes place in case of matching.
- ✓ A suitable substrate is applied to the buffer solution and it is enzymatically cleaved by the label.
- ✓ Resulting species starts an electrochemical redox process at the electrodes.
- ✓ Faradaic currents generated by the related redox process is detected and transduces DNA hybridization

# Enzymatically cleavage & redox process

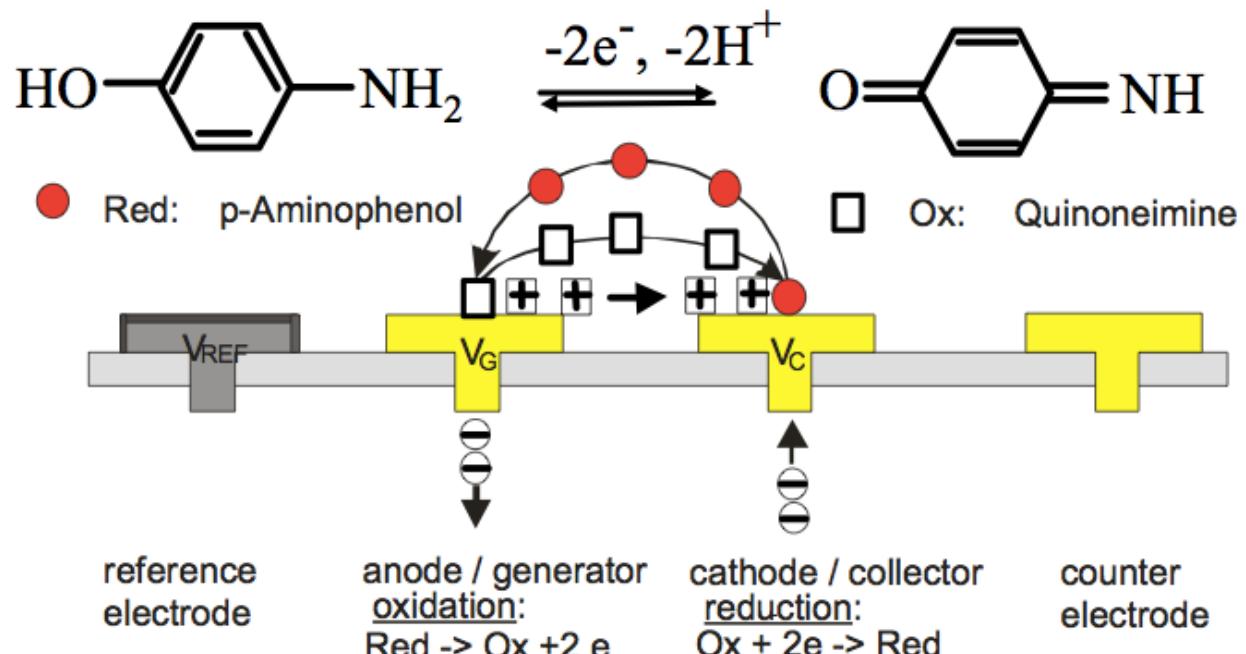
## a) Process at the label



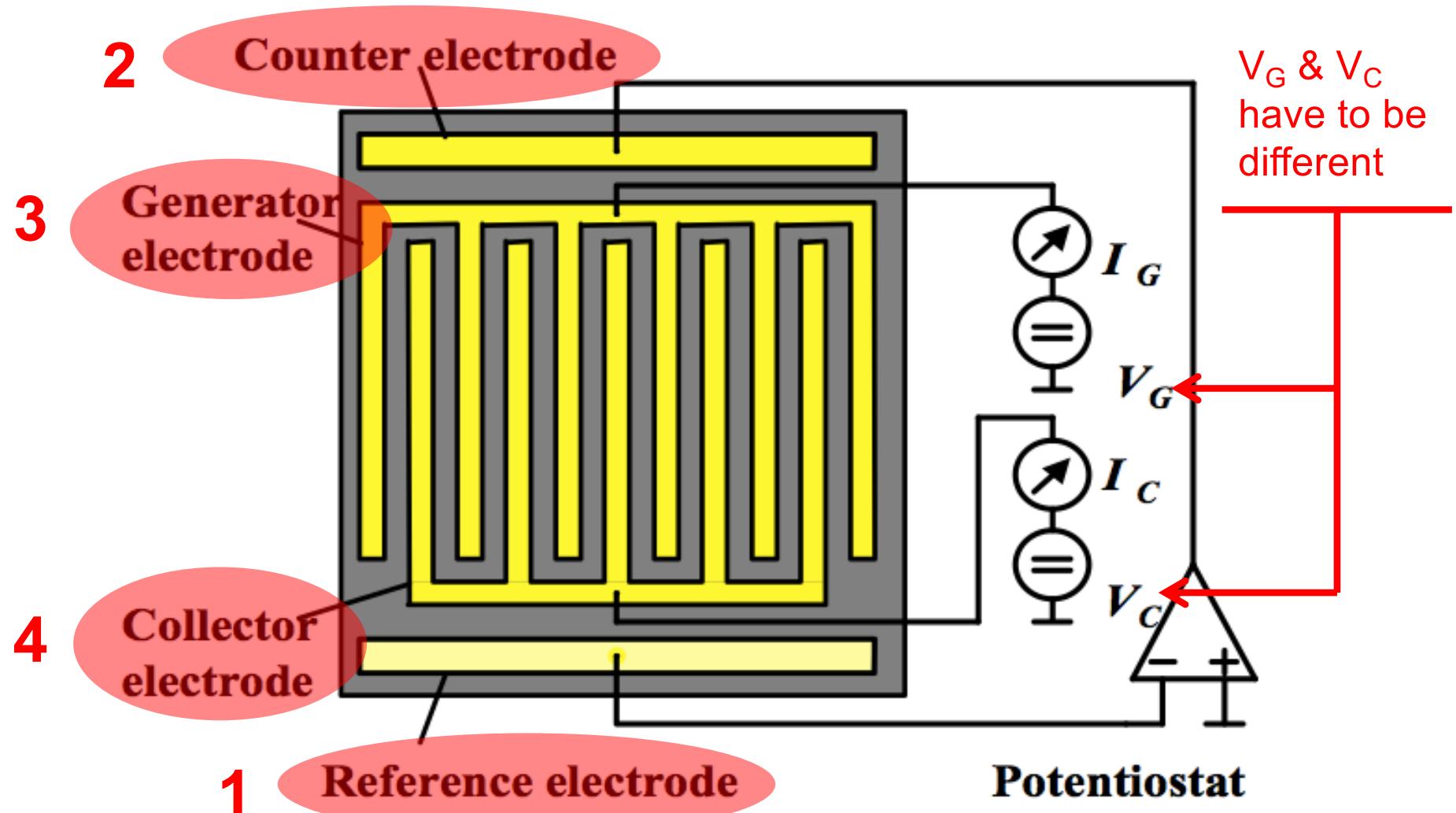
✓ The product of the cleavage is generating an oxidation process at the anode and, once oxidized, a reduction at the cathode

✓ The label cleaves the secondary probe

## b) Redox process at the electrodes

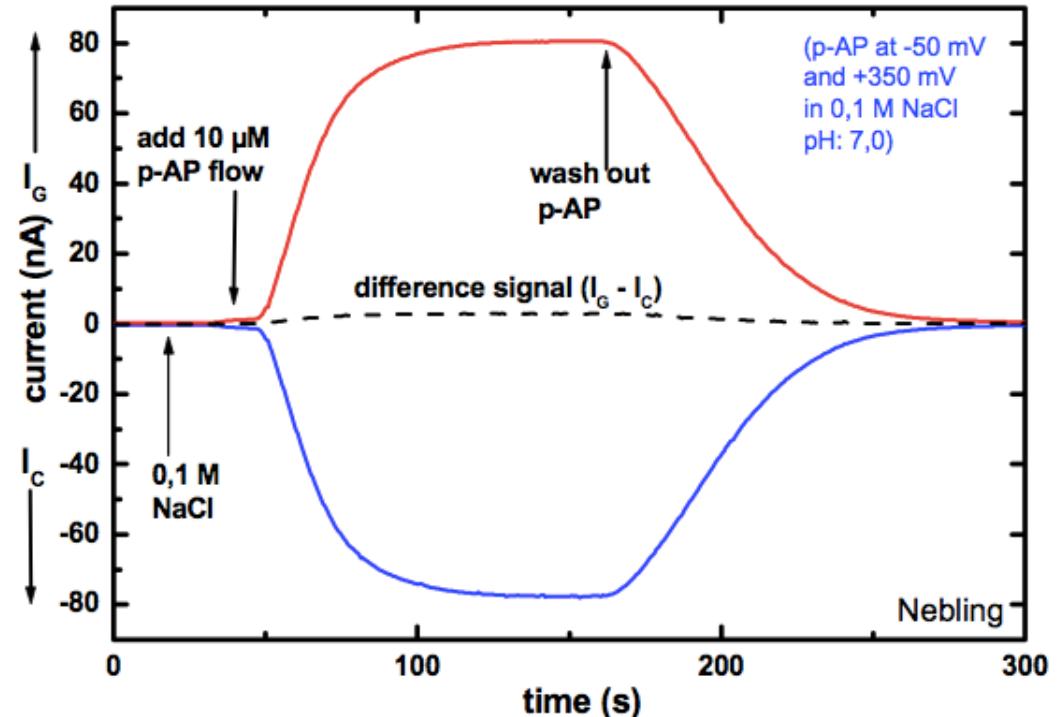
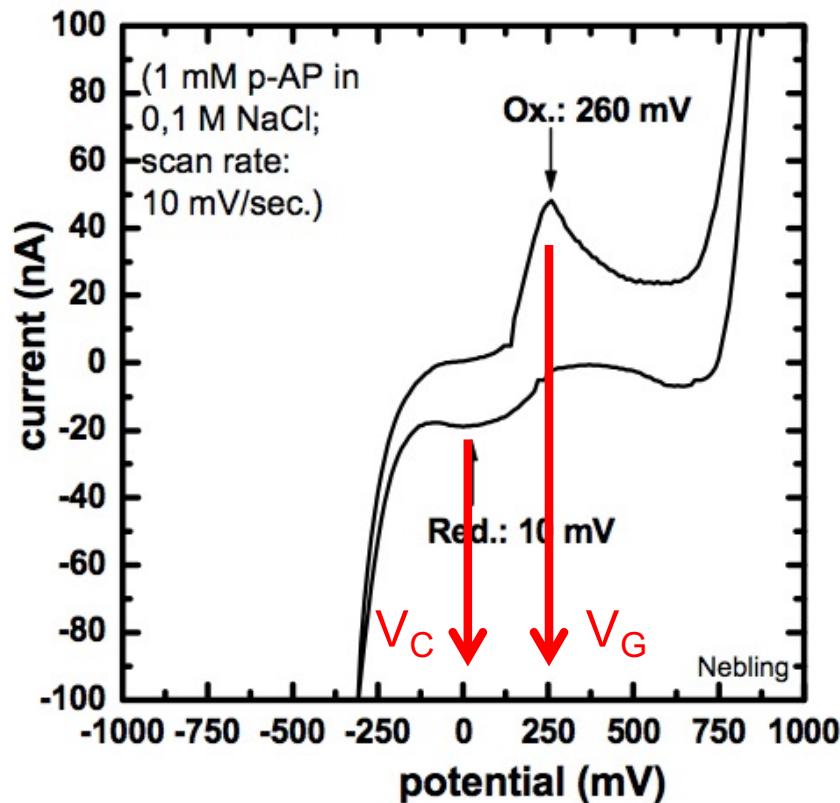


# The Electrochemical Cell



A 4-electrode Electrochemical Cell is here required

# Cyclic Voltammetry

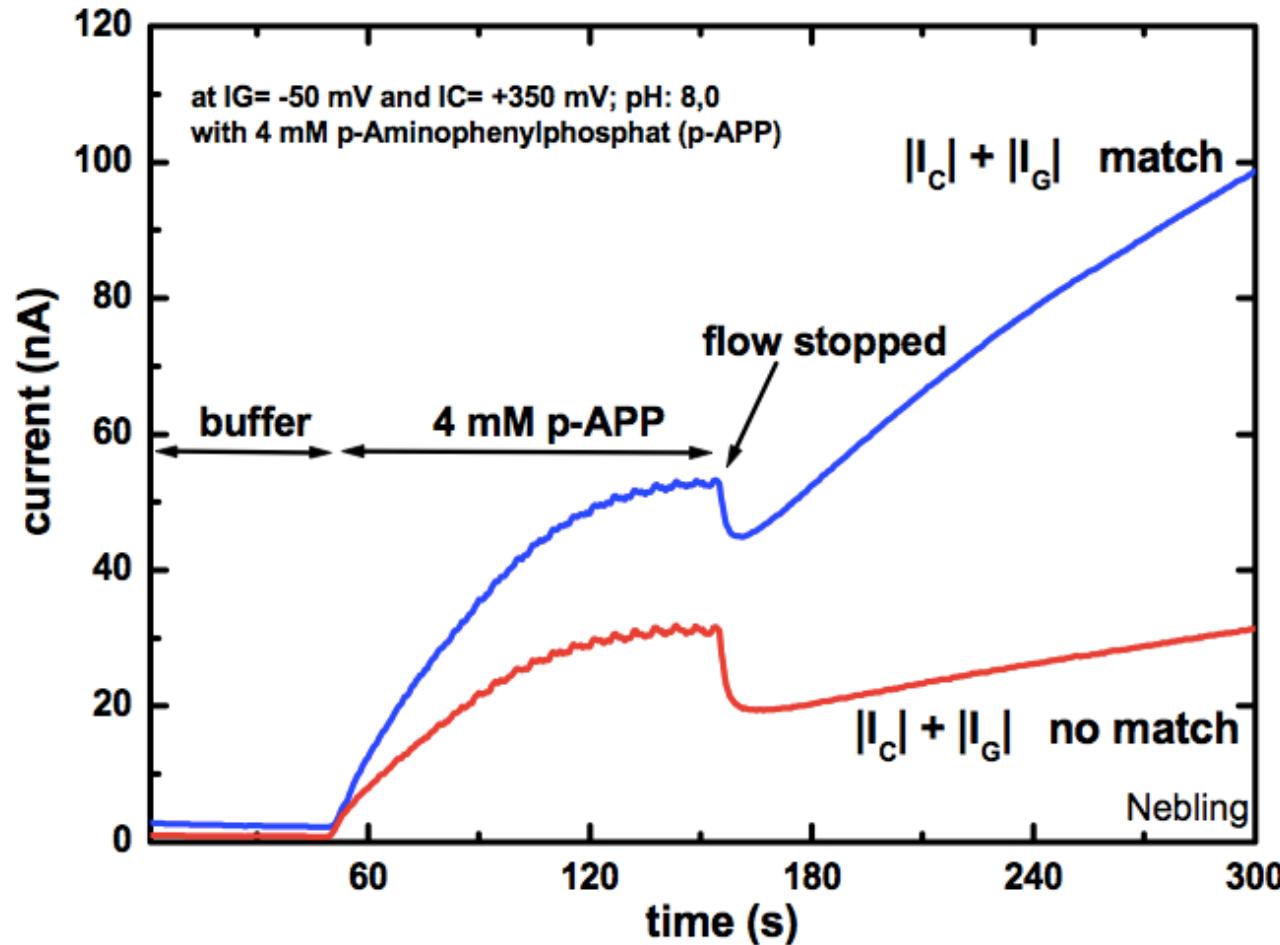


- ✓ Typical Cyclic Voltammetry acquired with 3-electrode cell

- ✓ Chronoamperometry acquired with 4-electrode cell

Simultaneous acquisition of Ox/Red current with 4-el

# Match/Mismatch DNA Hybridization



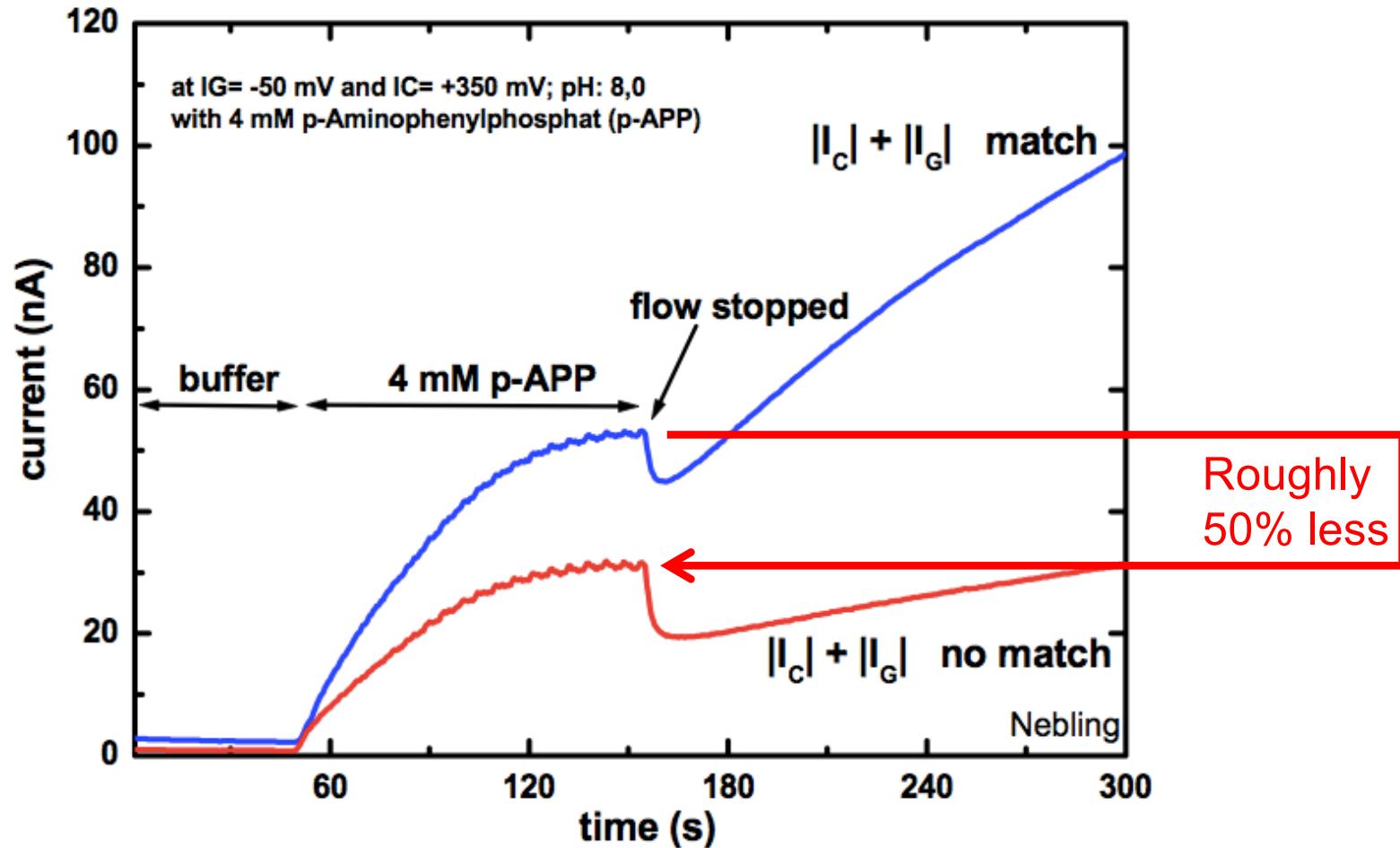
Successful detection of the matching sequence by significant signal by non-matching ones too

# Gibbs free energy for Match/Mismatch

duplex	<i>Experimental</i> $\Delta G$ [kJ/mol]
GGTTATTGG CCAATAACC	-26.8
GGTTCTTGG CCAAGAACCC	-31.4
GGTTTTGG CCAAAAAACC	-29.5
GGTTATTGG CCAAAAAACC	-12.0
GGTTCTTGG CCAATAACC	-12.4
GGTTTTGG CCAAGAACCC	-17.5

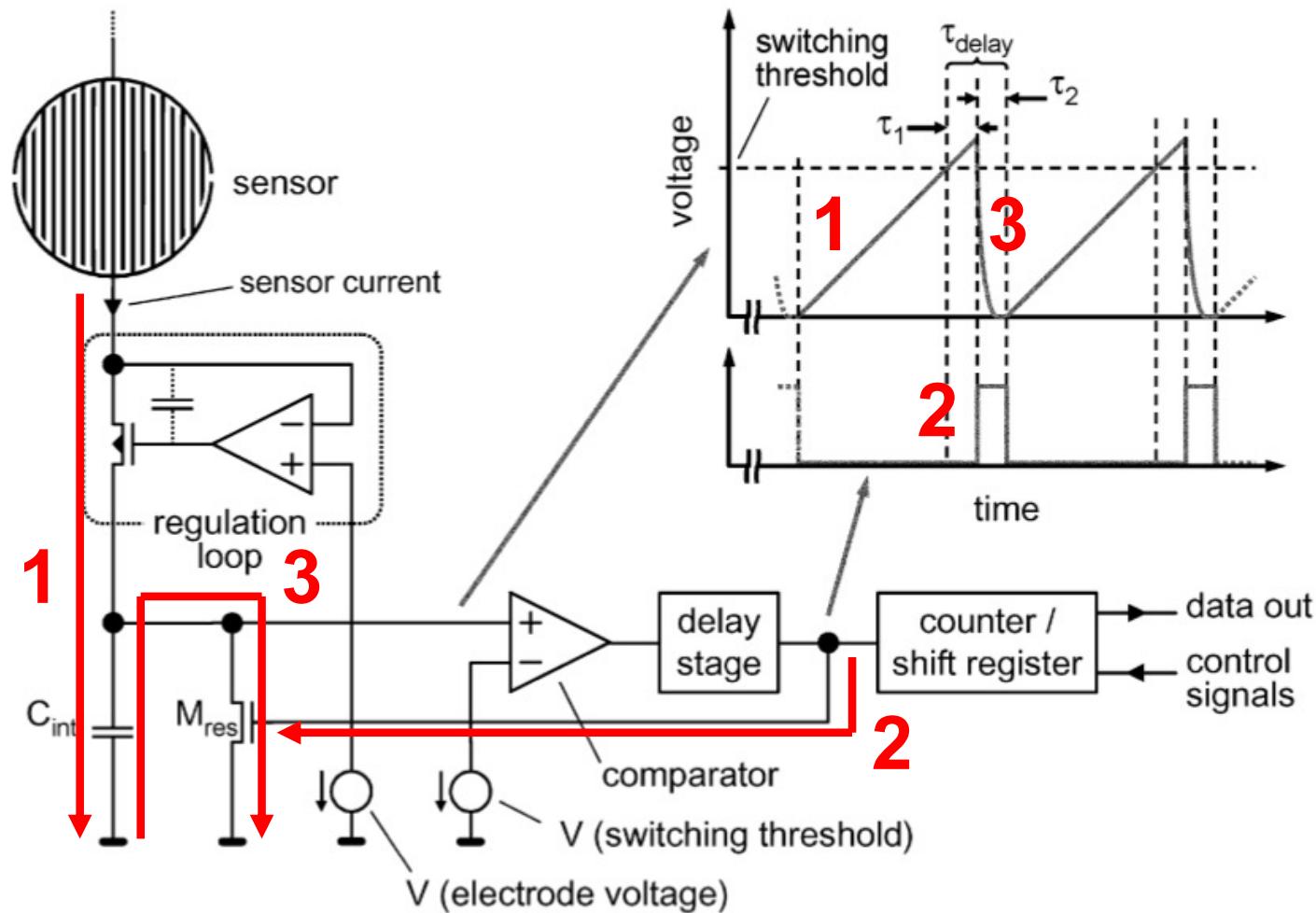
(c) S.Carrara

# Match/Mismatch DNA Hybridization



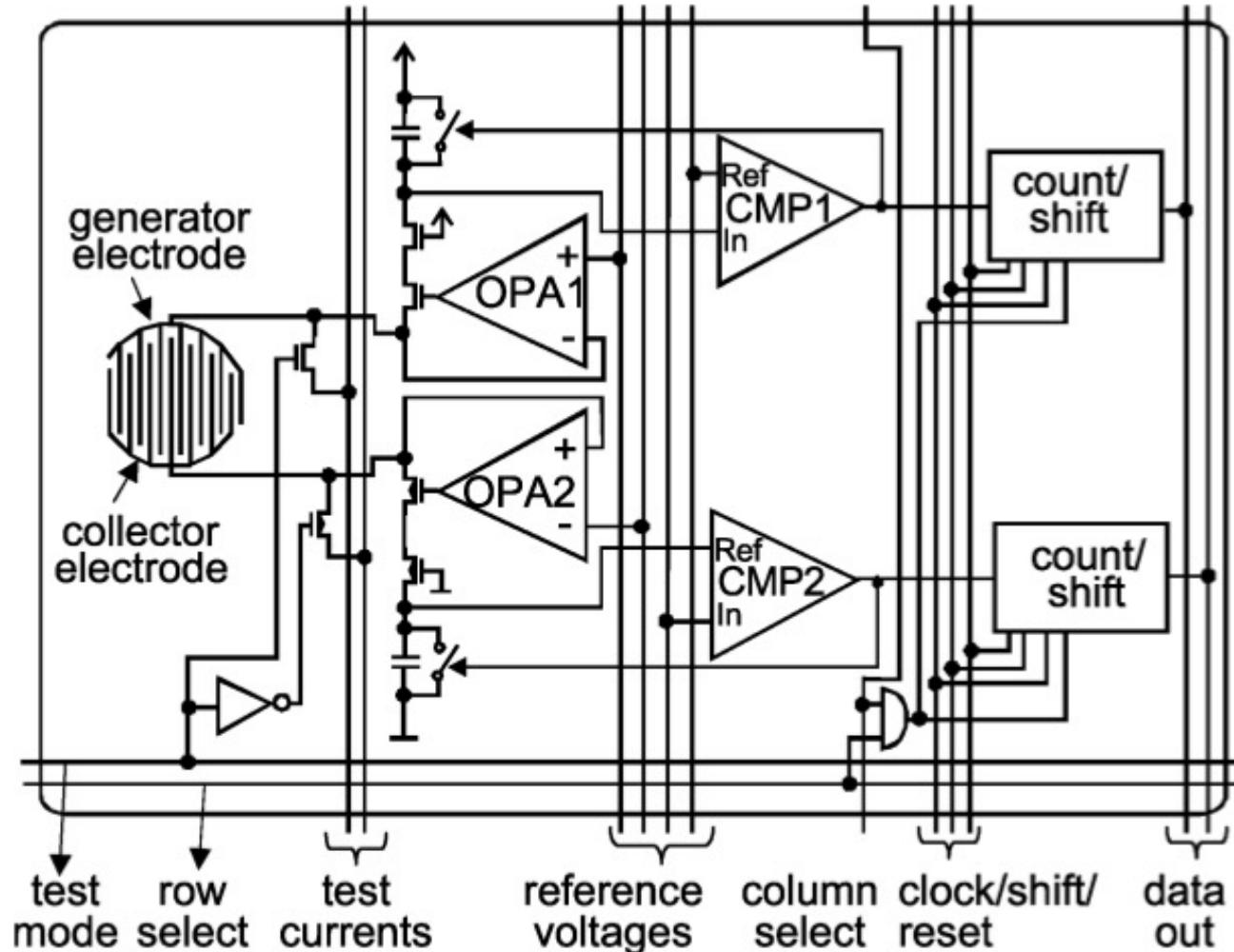
Successful detection of the matching sequences  
but significant signal by non-matching too

# Current CMOS Readout



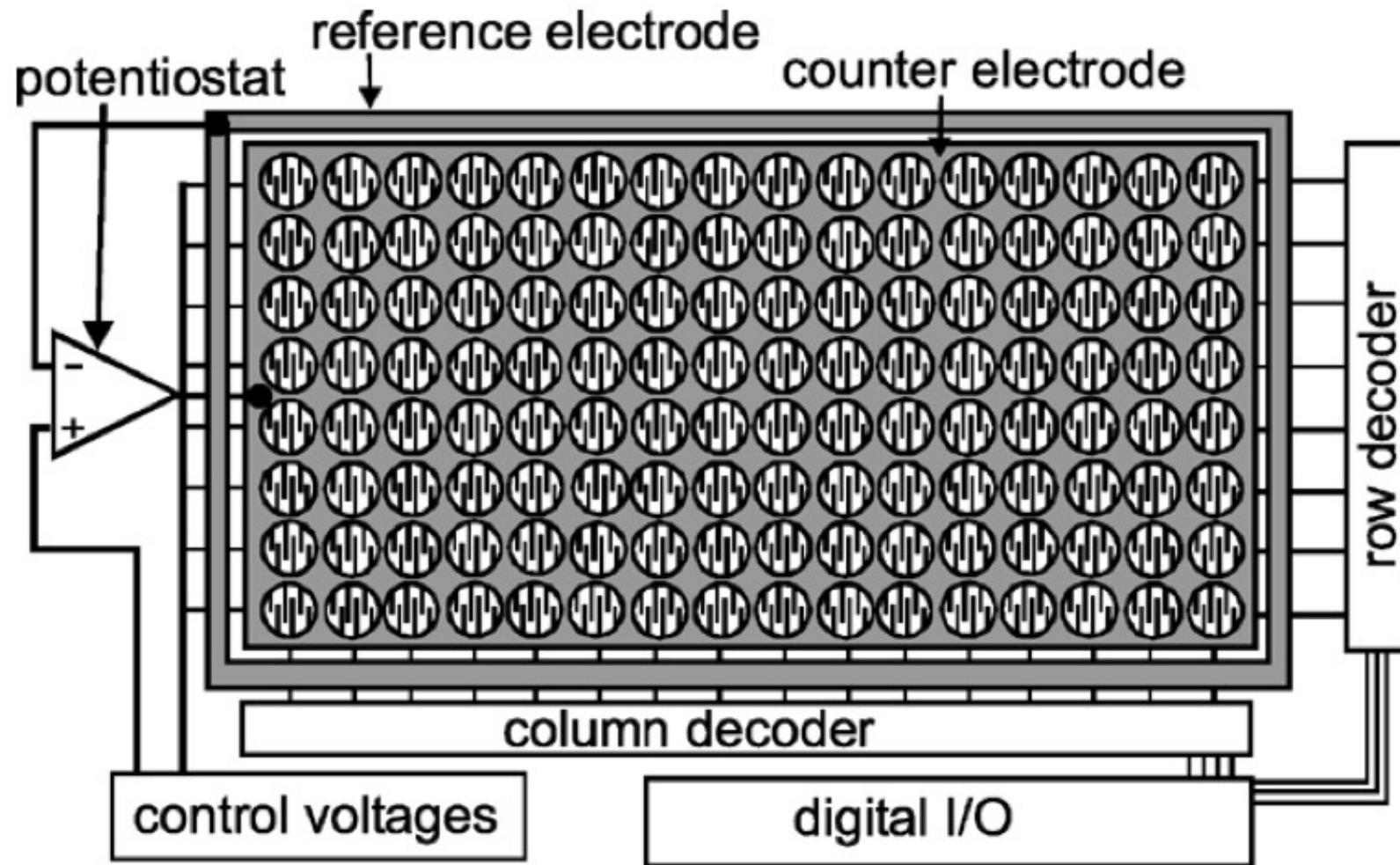
Frequency-To-Current Conversion (FTCC)  
method is used here too

# Current CMOS Readout



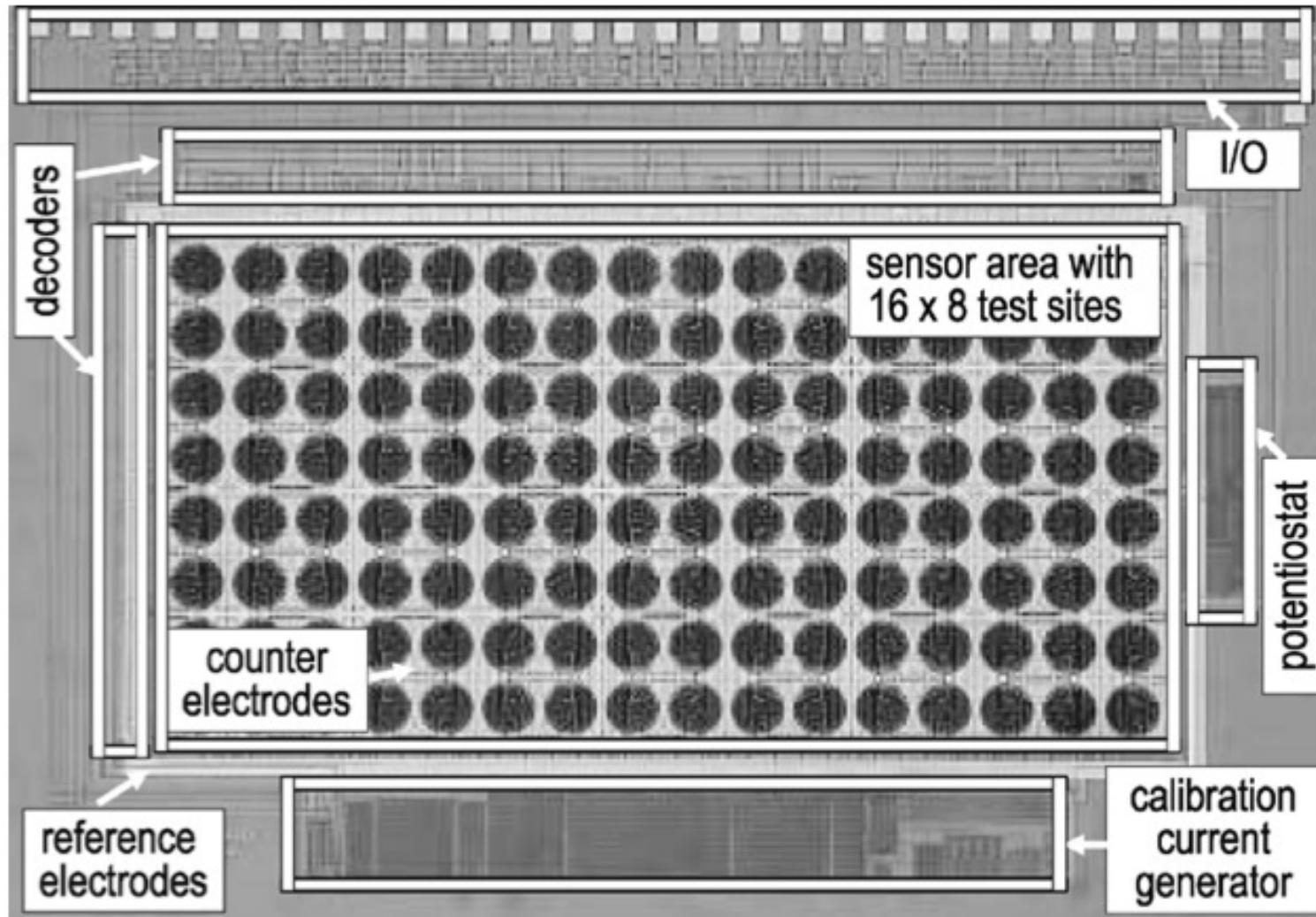
Sensor-site circuit architecture with digital output

# Array Architecture



Whole Chip architecture including Row/Column decoders

# The realized IC



Chip microphotograph. Total dimensions are  $6.4 \times 4.5 \text{ mm}^2$ .

# Exposed IC-Die Electrodes

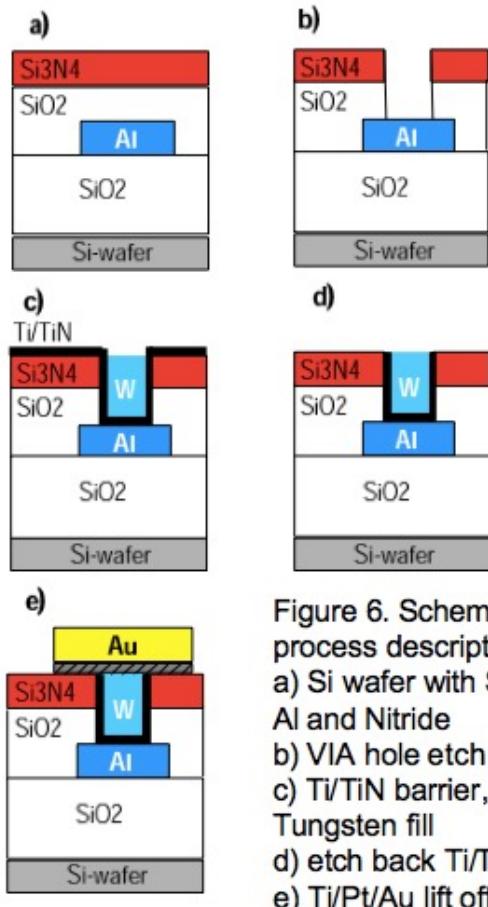
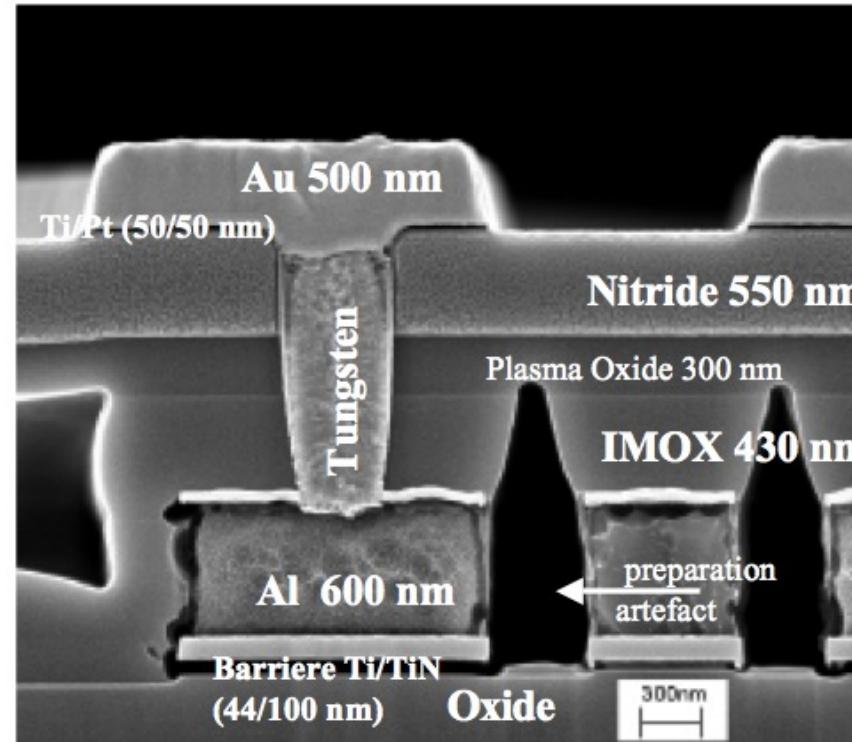
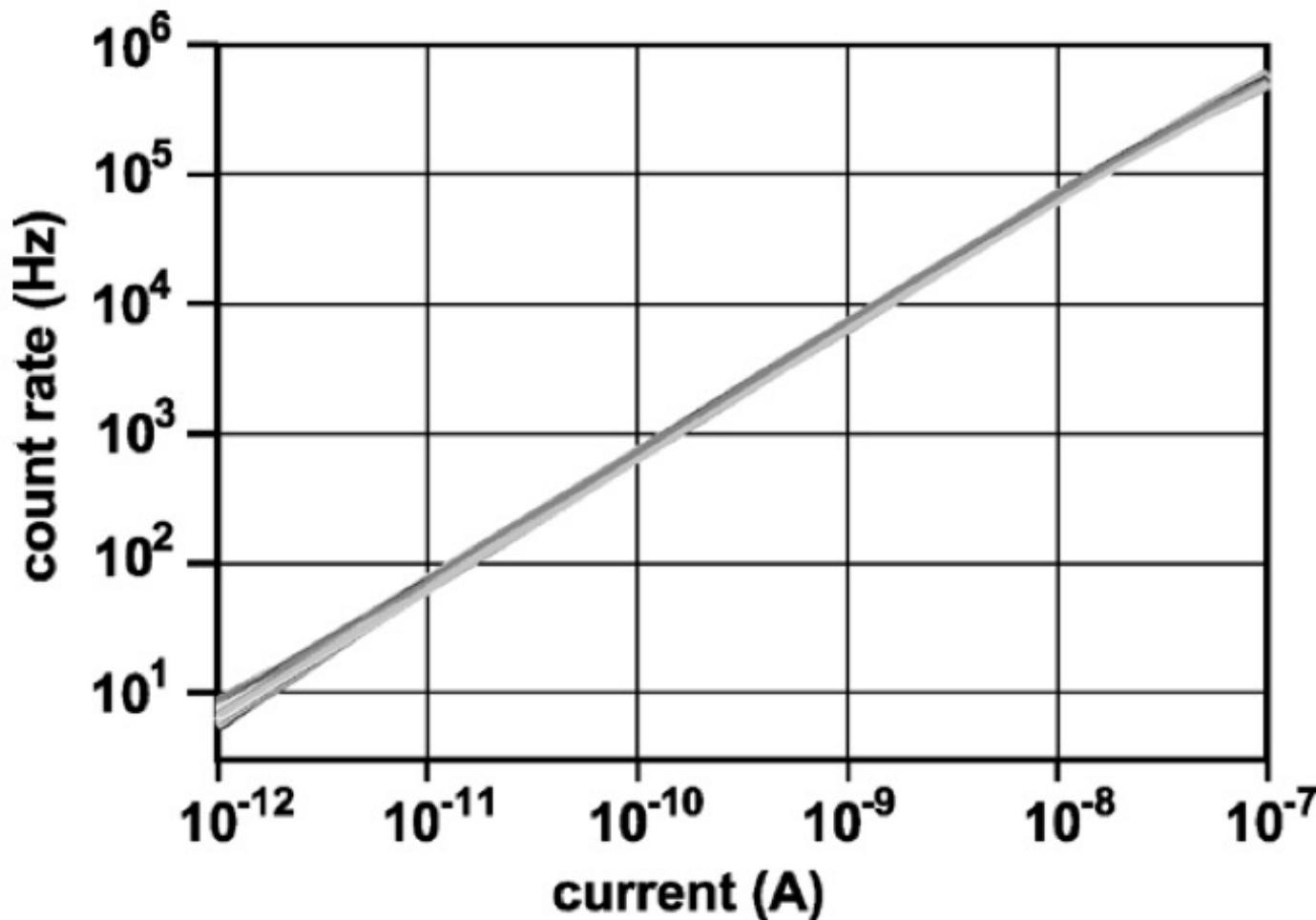


Figure 6. Schematic process description  
a) Si wafer with  $\text{SiO}_2$ , Al and Nitride  
b) VIA hole etch  
c) Ti/TiN barrier, Tungsten fill  
d) etch back Ti/TiN  
e) Ti/Pt/Au lift off



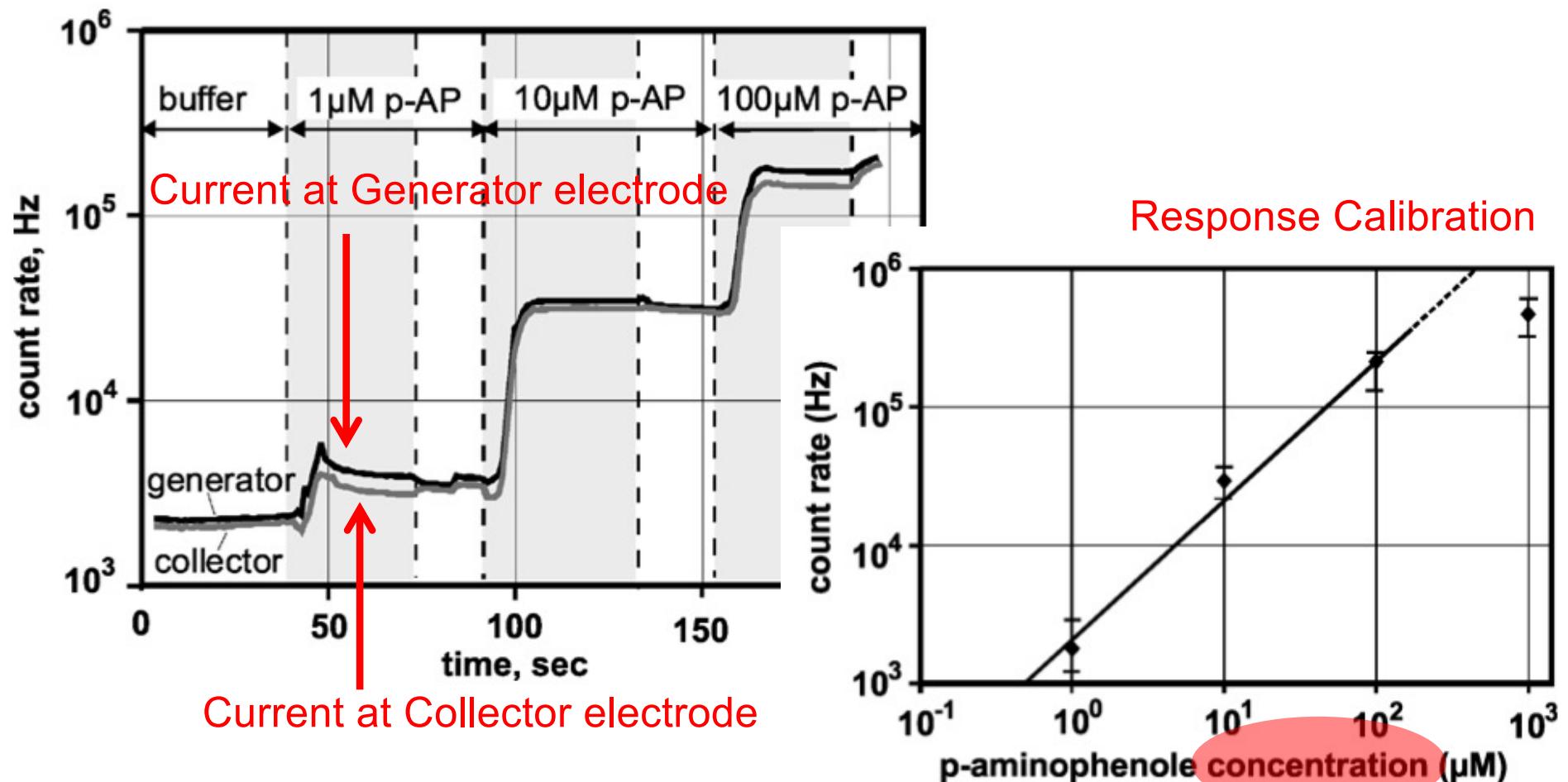
Electrochemical electrodes are created on top of the last CMOS metal Al layer

# Frequency Readout



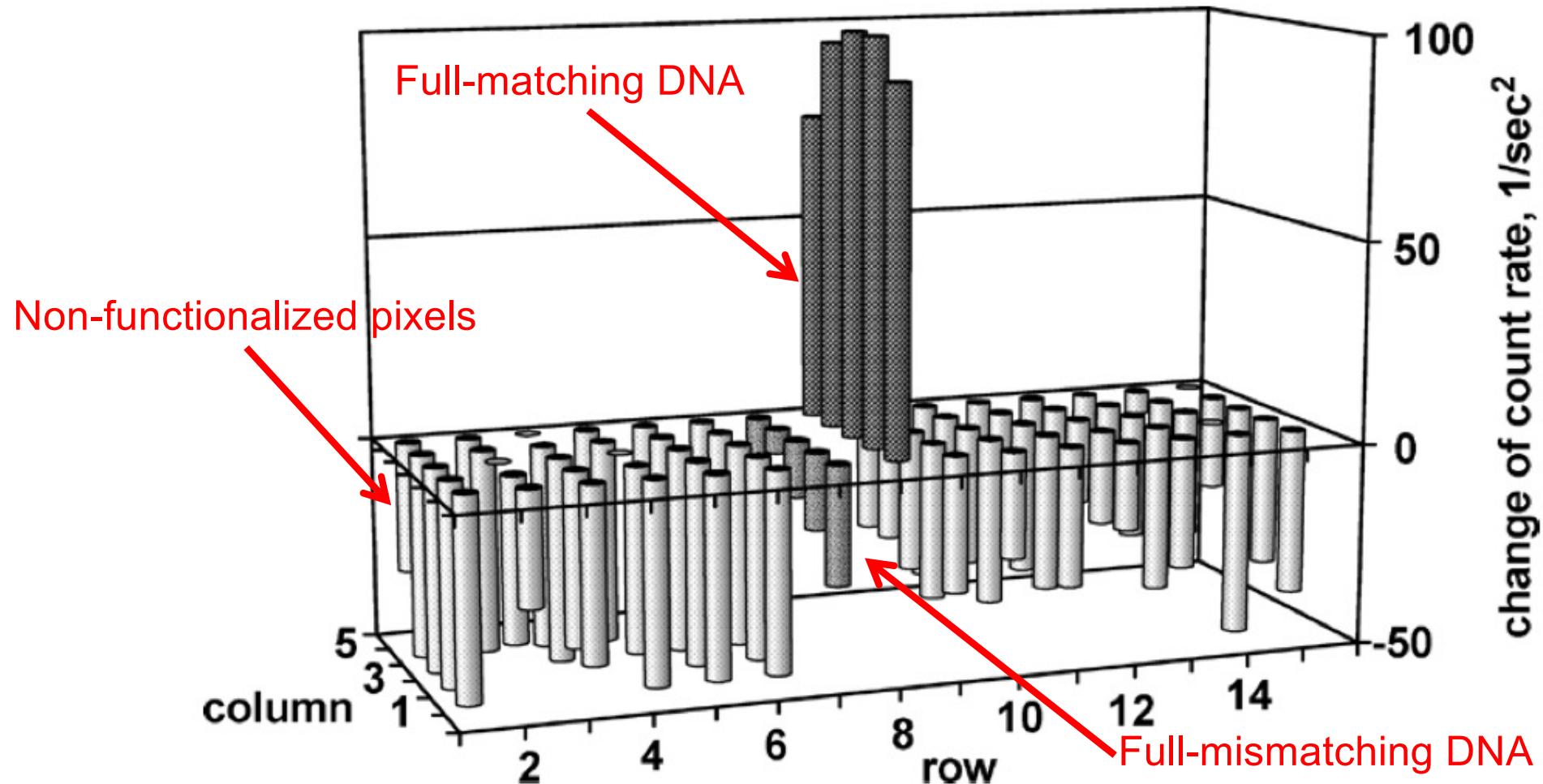
Measured count rate of all 128 DNA sensors

# Test Measures



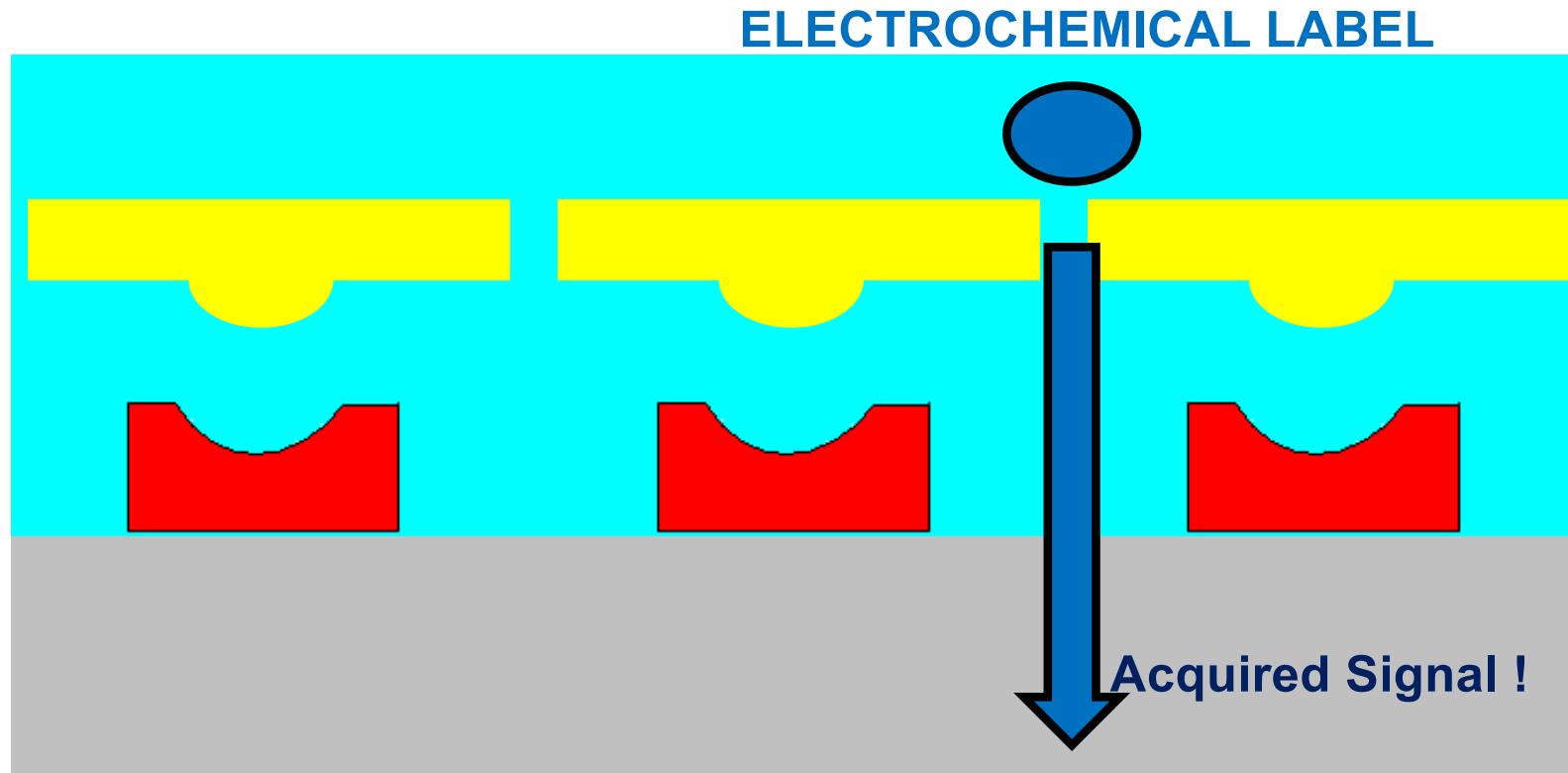
Response of the sensor versus secondary probe

# DNA Detection



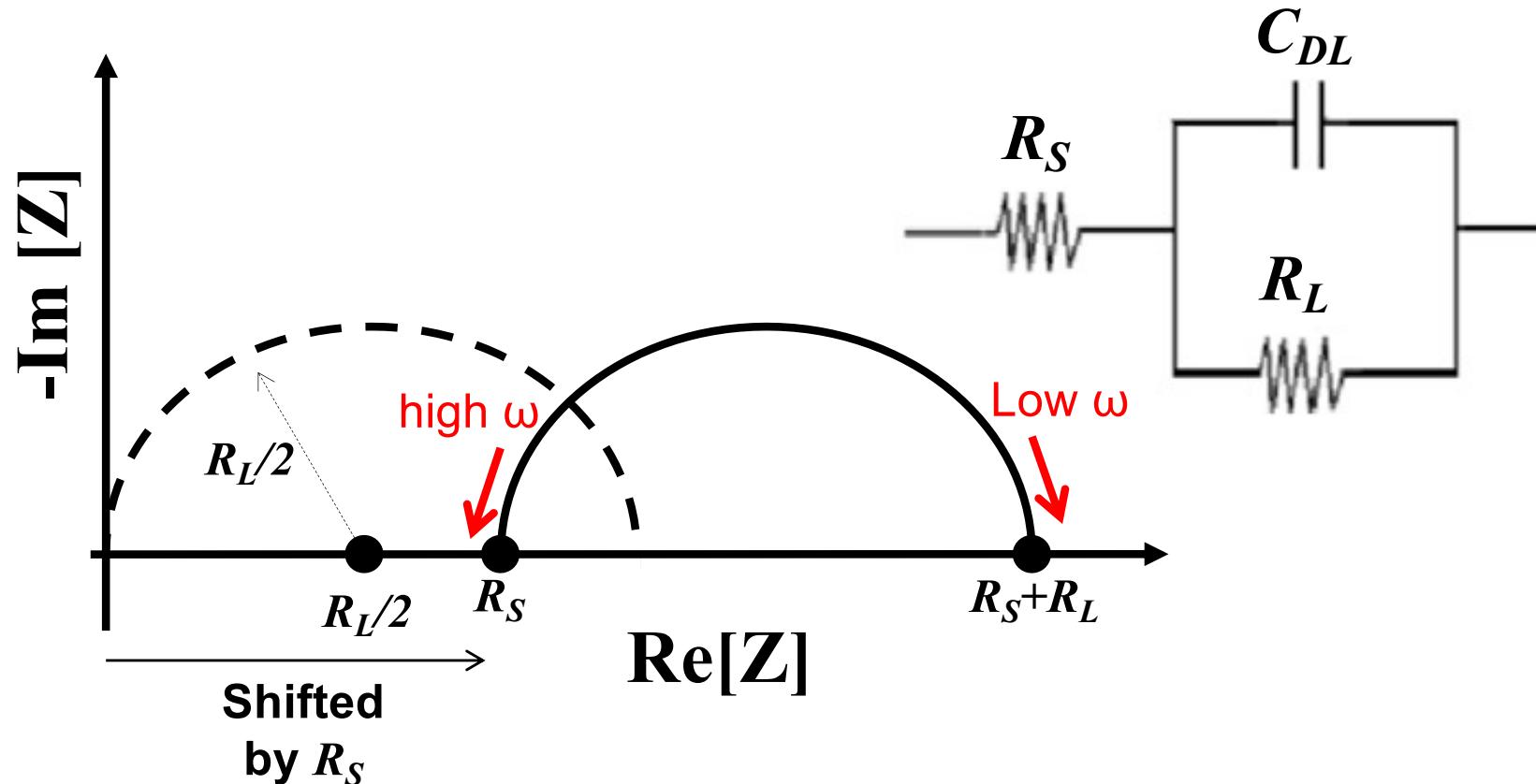
Row 8: full matching DNA, row 7: full mismatching DNA, all other positions not functionalized.

# Impedimetric Sensing



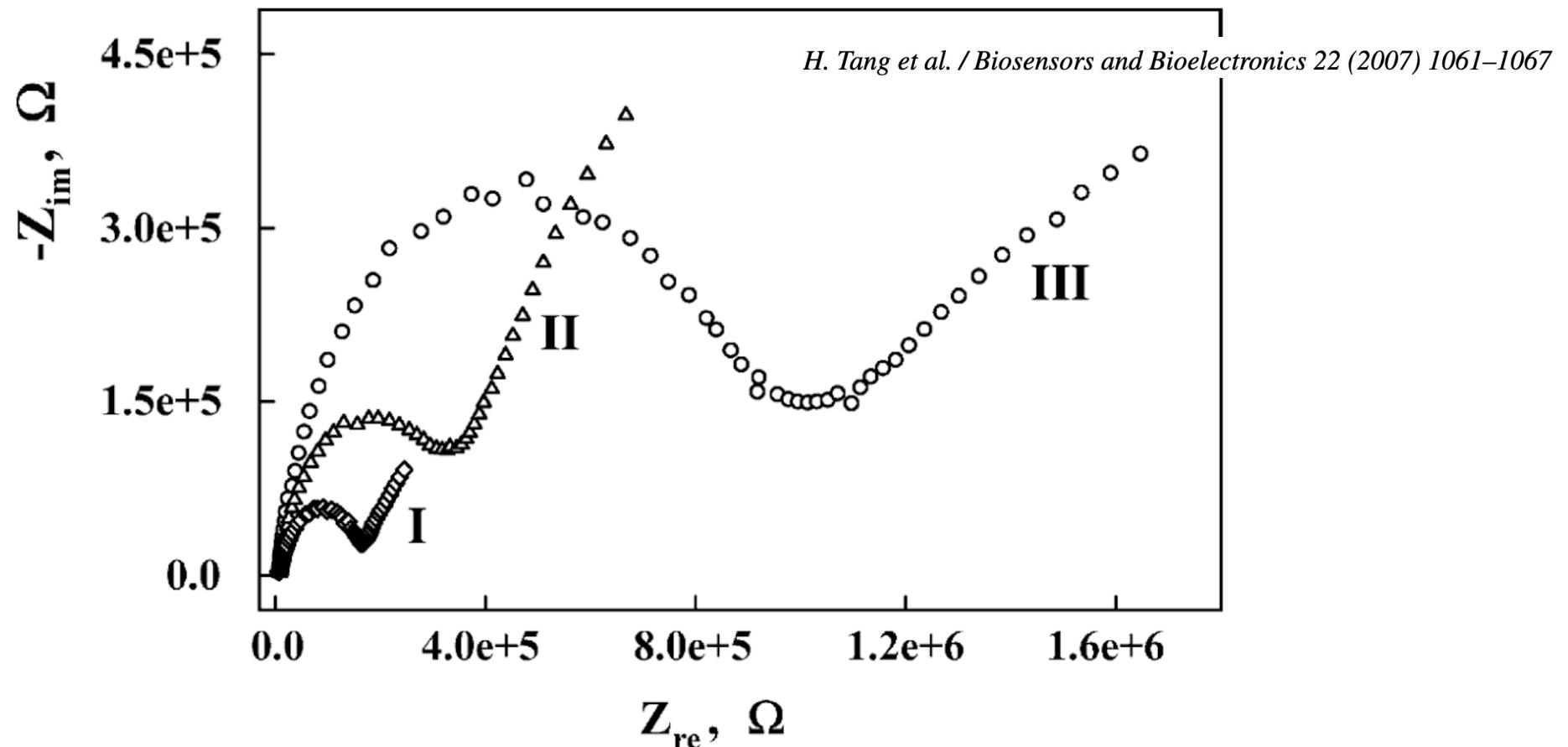
Clearly, the impedance of the Bio/CMOS interface changes accordingly to molecular steric hindrance

# Equivalent Impedance



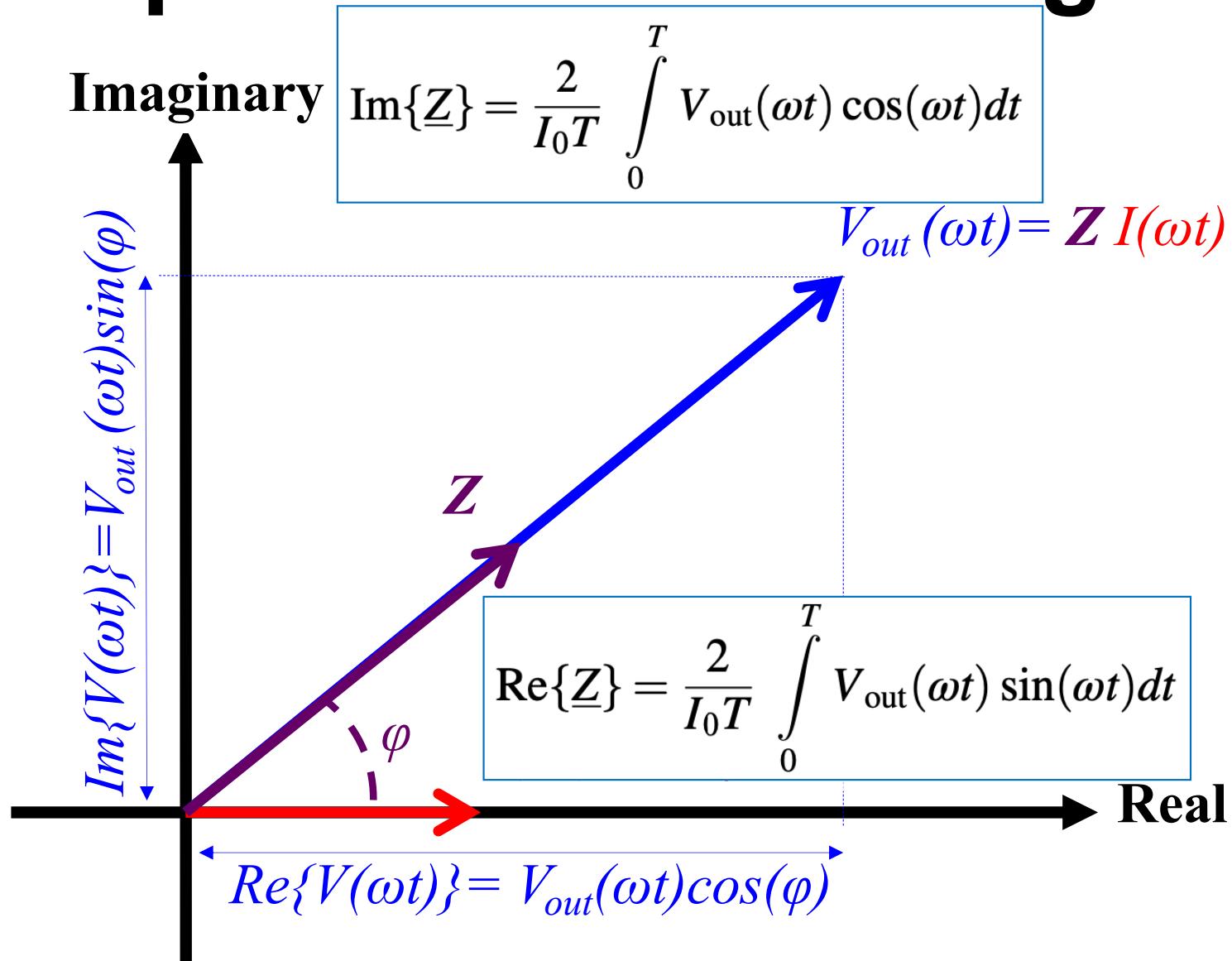
The Layering effects result in the impedance in parallel

# Impedimetric Sensing of CEA



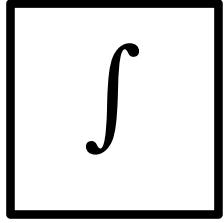
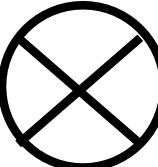
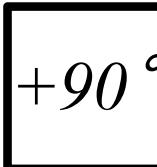
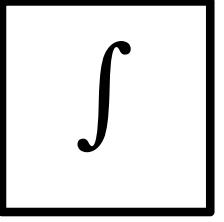
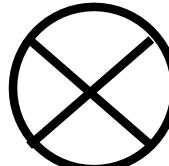
Nyquist plots: I-bare ; II-Antibodies ; and III-uptake with CEA (CarcinoEmbryonic Antigen)

# Impedimetric Sensing

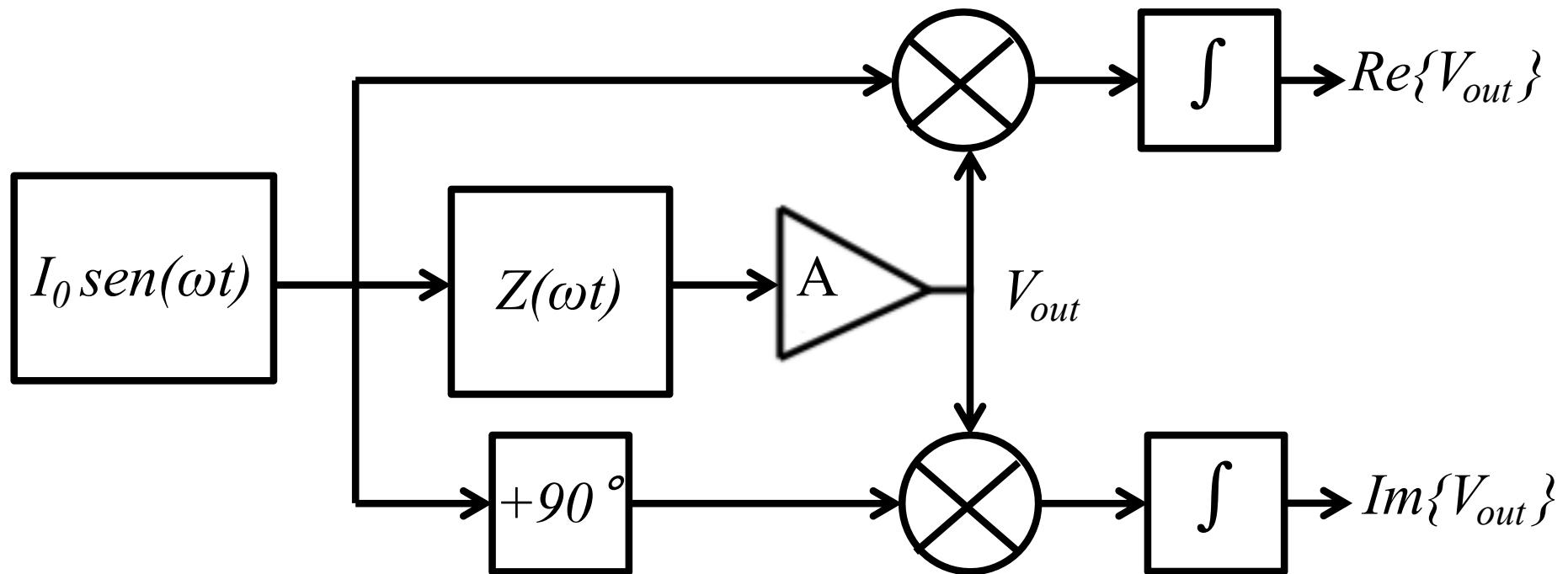


# Impedimetric Sensing

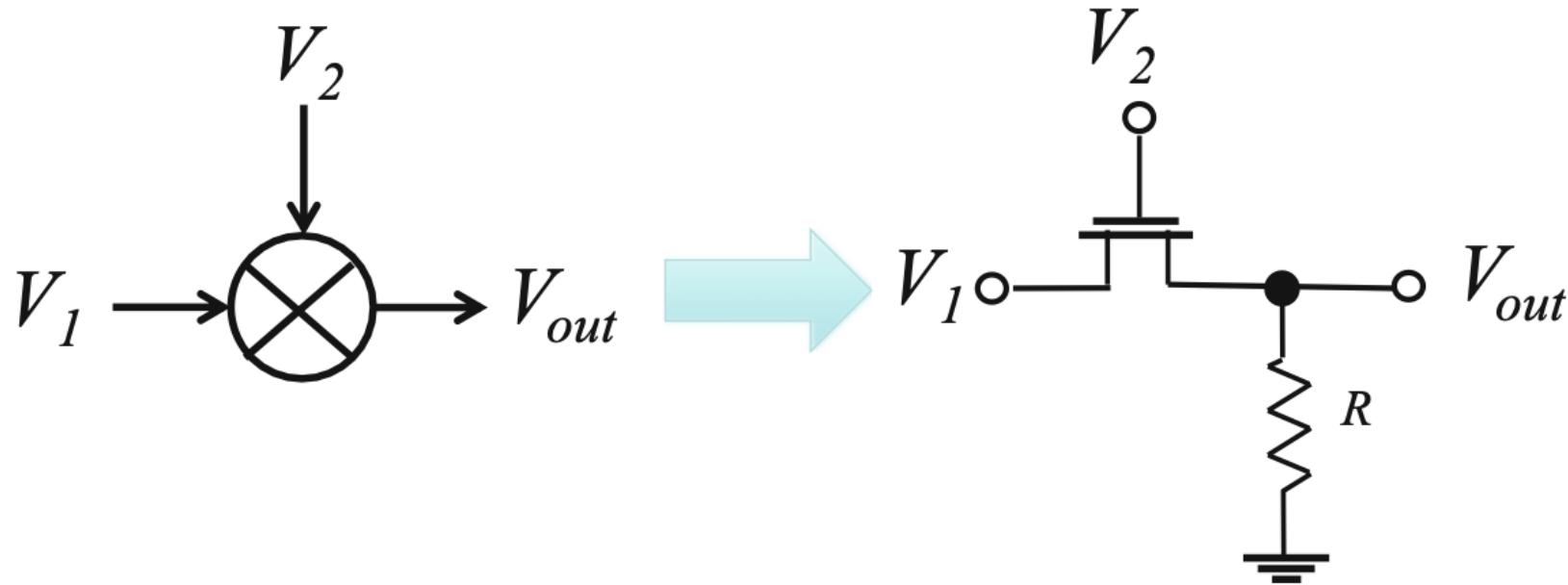
$$\left\{ \begin{array}{l} \text{Re}\{\underline{Z}\} = \frac{2}{I_0 T} \int_0^T V_{\text{out}}(\omega t) \sin(\omega t) dt \\ \text{Im}\{\underline{Z}\} = \frac{2}{I_0 T} \int_0^T V_{\text{out}}(\omega t) \cos(\omega t) dt \end{array} \right.$$

# Impedimetric Sensing

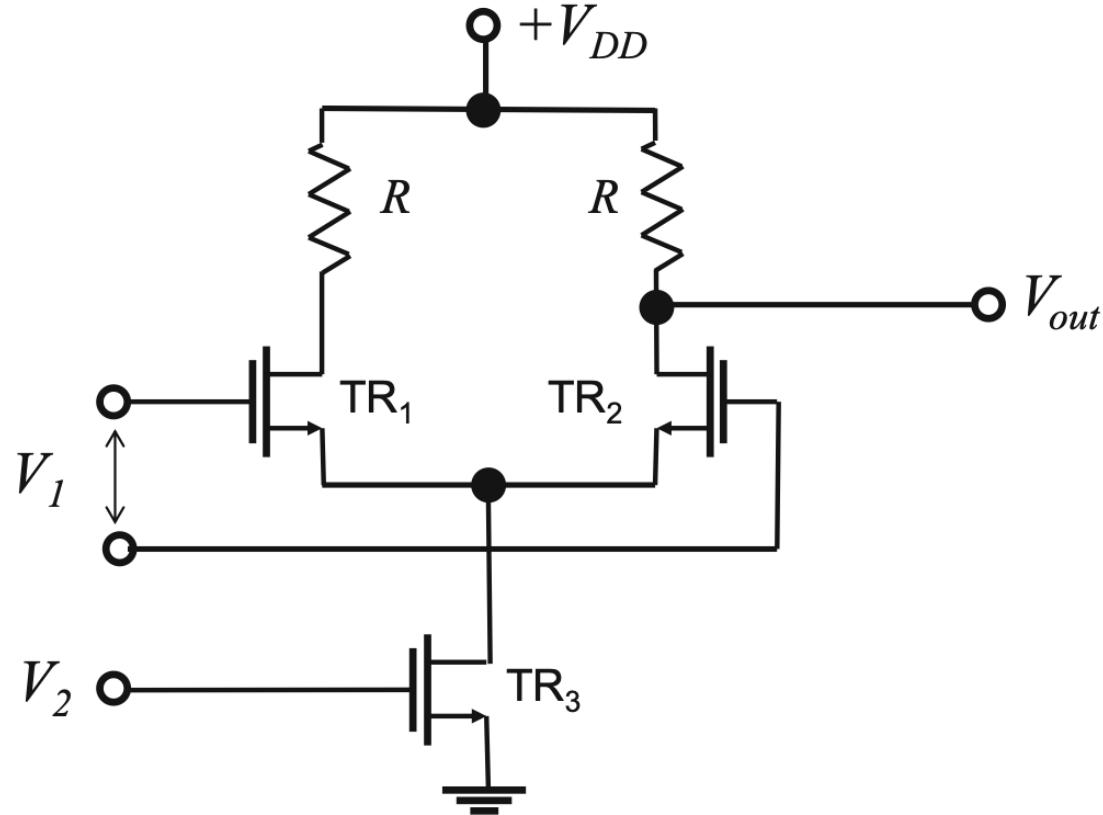


# Impedimetric Sensing



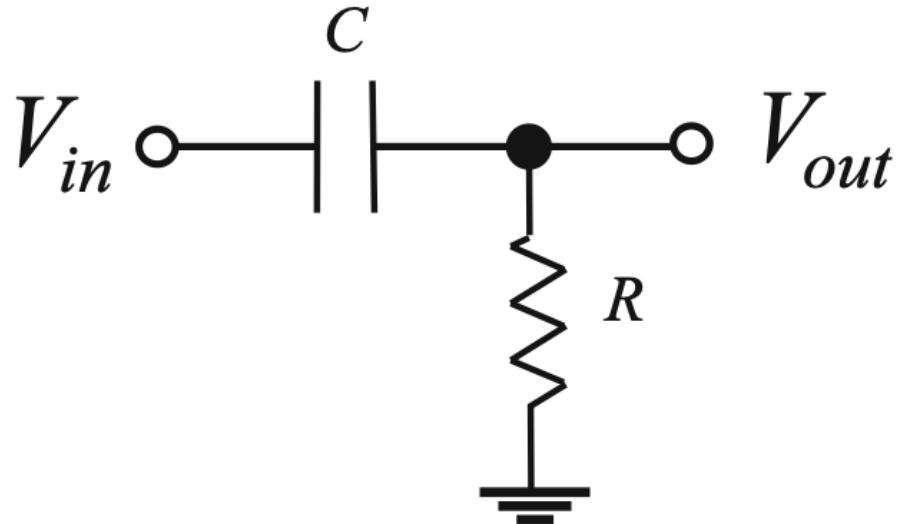
Voltage mixer and its simplest implementation with CMOS transistors

# Impedimetric Sensing



Voltage mixer implemented with  
a single-balanced mixer

# Impedimetric Sensing



Simplest possible phase shifter